# Vellus-to-terminal Hair Follicle Reconversion Occurs in Male Pattern Balding and is Promoted by Minoxidil and Platelet-rich Plasma: In Vivo Evidence from a New Humanized Mouse Model of Androgenetic Alopecia

Amos GILHAR<sup>10</sup>, Aviad KEREN<sup>1</sup> and Ralf PAUS<sup>2-4</sup>

<sup>1</sup>Skin Research Laboratory, Rappaport Faculty of Medicine, Technion – Israel Institute of Technology, Haifa, Israel, <sup>2</sup>Dr Phillip Frost Department of Dermatology & Cutaneous Surgery, Miller School of Medicine, University of Miami, Miami, FL, USA, <sup>3</sup>Monasterium Laboratory, Münster, Germany and <sup>4</sup>CUTANEON, Hamburg, Germany. E-mail: doritg2000@gmail.com Accepted Aug 15, 2023; Published Oct 18, 2023

Acta Derm Venereol 2023; 103: adv12320. DOI: 10.2340/actadv.v103.12320

It has recently been claimed that, once terminal (T) scalp hair follicles (HFs) have been fully miniaturized into vellus (V) HFs in male pattern androgenetic alopecia (mpAGA), these cannot be reconverted into T HFs, even under therapy (1), and that even long-term treatment with topical minoxidil 5% (MXL) or oral finasteride fails to significantly change the number of V HFs (1). Rushton & Van Neste et al. (1) reported that hair regrowth following treatment for AGA is mostly due to an increase of HFs in kenogen, i.e. when the HF does not contain a visible hair shaft (2). Instead, these authors postulated that T HFs in a period of relative dormancy are re-activated by mpAGA therapeutics and that any increase in the number of hair shafts observed post-therapy primarily reflects anagen initiation in kenogen HFs (1).

This provocative concept stands in sharp contrast to prior histological evidence (3), and would have major clinical consequences: it would mandate early intervention in AGA management after which only hair transplantation and, perhaps, selected cell-based therapies would be therapeutically meaningful (4, 5).

Yet, the fact that hypertrichosis-inducing drugs (e.g. MXL, cyclosporine A) and hormones (e.g. androgens, adrenocorticotropic hormone, and even cortisol) can quite rapidly convert V into T HFs (6) questions the non-convertibility hypothesis. The latter is based on unit area trichogram and phototrichogram, which relies on defining V HFs based on their visible hair shaft and thus has inherent methodological limitations (7).

## **MATERIALS AND METHODS**

Quantitative histomorphometry of horizontally sectioned lesional mpAGA scalp skin biopsies are the most accurate method for assessing HF miniaturization and its reversal (3, 8, 9). Using this more accurate methodology, we asked in the current pilot studywhether a former T HF, which has been transformed into a V HF under androgen-stimulation, can be reconverted to a T HF under standard AGA therapy in a well-defined area of mpAGA scalp skin in vivo. To address this question, we have re-analysed human scalp skin samples derived from our novel humanized mpAGA mouse model, in which human mpAGA scalp xenotransplants had been treated long-term with MXL or platelet-rich plasma (PRP) in vivo (10, 11), applying the objective histomorphometry criteria listed in Fig. 1A.

A total of 57 lesional biopsies from 10 mpAGA patients (mean age  $35.9 \pm 9.4$  years) obtained after informed patient consent and IRB approval, which had been transplanted onto SCID/beige mice (10, 11), were re-examined histologically. Five mice were treated

once daily with 5% topical MXL. 4 mice with vehicle. 5 mice once monthly for 4 months with intradermal injection of non-activated autologous PRP (control) and 5 mice with activated autologous PRP, prepared and activated as described (10). Each mouse was transplanted with 3 xenotransplants. In our previous reports (10, 11), we had not interrogated therapy-induced changes in the % of V, "intermediate", and T HFs in the mpAGA-affected human scalp skin xenotransplants. Namely, we searched for post-therapy changes in the number of V HFs with an associated arrector pili muscle (APM), whose presence is thought to identify those V HFs that once had been a T HF (7), while HFs that always were V typically lack an APM (12).

Quantitative histomorphometry of the xenotransplants after 4 months of therapy with either MXL or PRP compared with vehicle or non-activated-PRP showed a significantly decreased number of V HFs with an associated APM (p < 0.05), i.e. of those HFs that likely had undergone prior  $T \rightarrow V$  conversion during mpAGA (7) (Fig. 1B and Tables SI and SII). Moreover, compared to vehicletreated controls, the MXL- or PPR-treated mpAGA scalp skin xenotransplants also showed significantly fewer intermediate HFs (p < 0.01) (Fig. 1C, D and Tables SI and SII). Intermediate HFs are overall shorter, and display smaller hair shaft, bulb, and dermal papilla diameter, and thus represent a  $T \rightarrow V$  transition stage that precedes complete HF miniaturization (4, 13). Inversely, this was accompanied by a significant increase in the number of T HFs (p < 0.05) in MXL- or PPR-treated xenotransplants compared with vehicle or non-activated PRP (Fig. 1D).

The number of V without APM remained constant between the test and control xenotransplant groups (Fig. 1D). This is interesting in the context of Sinclair's hypothesis (7) that loss of contact between the APM and the stem cell-rich bulge in AGA may render HF miniaturization irreversible (14) and might be interpreted as supporting this hypothesis. Alternatively, this could indicate that HFs in mature adult human scalp skin, which always represented the V phenotype, are much less responsive to MXL and PRP treatment than V HF that are miniaturized former T HFs.

#### DISCUSSION

These histomorphometric data demonstrate that, in line with Whiting (3),  $V \rightarrow T$  reconversion of V HFs can indeed occur under therapy, even in long-standing mpAGA, at least in this humanized mouse model in vivo. Theoretically the reduced number of vellus HFs after therapy may also reflect deletion of vellus HFs, as HFs can undergo "programmed organ deletion" (15), although no AGA therapy has been shown to promote this phenomenon.

Of course, one needs to consider that xenotransplantation is initially associated with a temporary wound-healing response and that male mouse testosterone serum levels tend to be substantially lo-

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**Fig. 1.** Arrector pili muscle in male pattern androgenetic alopecia (mpAGA) xenotransplants treated with activated platelet-rich plasma (PRP) or minoxidil 5% (MXL) vs control. (A) Overview of the experimental design. Biopsies were obtained from 10 patients with androgenetic alopecia (AGA) and transplanted onto 19 severe combined immunodeficiency (SCID)/beige mice, which were treated with either MXL or PRP (10, 11). The paraffin-embedded sections were cut and stained with haematoxylin and eosin (H&E) to define the hair follicles (HFs) according to their hair shaft diameter (3) and with Mason's trichrome to distinguish the red-staining arrector pili muscle (APM) (7). (B) Higher magnification of vellus (V) HFs with preserved APM in treated vs vehicle control xenotransplants. (C) Representative photomicrographs of horizontal sections shows a predominance of V and intermediate HFs in mpAGA skin before transplantation compared with a majority of terminal (T) HFs in xenotransplants treated for 4 months with topical MXL. (D) Quantitative histomorphometry demonstrates that the number of V HFs without APM remained almost constant in treated vs control xenotransplants. In parallel, the number of T HFs in xenotransplants treated with PP or MXL increased significantly compared with the control xenotransplants. Data are presented as the mean ± standard error of mean (SEM). Statistical significance was set at *p*-value < 0.05 and calculated by non-parametric Kruskal-Wallis test, followed by a Mann–Whitney *U* test. *Scale bars:* 50 µm.

wer than those in human males, which might have impacted our results. However, murine dihydrotestosterone serum levels are actually quite similar to the human ones (16) which renders it unlikely that human scalp skin xenotransplants grow in a state of relative systemic hypoandrogenism thatmight have artificially facilitated V $\rightarrow$ T HF reconversion in our *in vivo* system. Moreover, potential HF reconversion effects of the initial wound-healing response post-transplantation would have affected both test and control xenotransplants and would hardly still be visible 4 months later.

Therefore, our pilot *in vivo* study questions the validity of the Rushton and Van Neste et al. (1) hypothesis by showing that standard clinical mpAGA therapy can promote a V $\rightarrow$ T reconversion in lesional human mpAGA skin in principle, at least in miniaturized V HFs that have not lost contact with their APM. While the current study data in a humanized mouse model of AGA obviously require biopsy- and histomorphometry-based confirmation in mpAGA patients under therapy, there is no biologically compelling reason why V $\rightarrow$ T HF reconversion should not also be possible under clinical conditions. As predicted from previous *ex vivo* work with organ-cultured human scalp HFs (4), the current study data also suggest that, unsurprisingly, intermediate HFs are more reconversion-responsive than fully miniaturized V mpAGA HFs (Fig. 1A, D and Tables SI and SII).

A long-term, large-scale, biopsy-based clinical study on patients with mpAGA, using the same definitive quantitative histomorphometry read-out parameters as employed here, is needed to confirm the findings of the current *in vivo* pilot study. It is possible that a larger clinical study could render non-surgical therapies in mpAGA management more promising than has recently been claimed (1), and make early therapeutic intervention strongly advisable.

#### ACKNOWLEDGEMENTS

This study was supported in part by the Technion Research and Development Foundation (TRDF) to A.G. and an Endowed Frost Scholarship to R.P.

*Conflicts of interest:* AG, AK and RP state no conflict of interest. For the record, in their laboratories AG and RP perform contract preclinical hair research for competing industry clients, and RP is founder & CEO of Monasterium Laboratory and CUTANEON, Germany.

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