

Endocrine regulation of craniofacial growth

Sinikka Pirinen

Institute of Dentistry, Department of Pedodontics and Orthodontics, University of Helsinki, Helsinki, Finland

Pirinen S. Endocrine regulation of craniofacial growth. *Acta Odontol Scand* 1995;53:179–185. Oslo. ISSN 0001-6357.

This paper presents an updated review of the role of endocrine factors in craniofacial and dental development. Some unpublished results of the author's own studies are presented. Longitudinal growth studies have shown the similarity of facial and somatic growth rates, whereas dental development has been found to be independent. The increased therapeutic use of the growth hormone (GH) has focused attention on the dental and craniofacial effects of GH. Whereas delayed and advanced cranial base and facial growth is obvious in conditions with either lack or excess of GH, the effect of GH on the dentition is less clear. Thyroid hormones seem to be necessary for the eruptive movement of teeth. Sex steroids clearly have an effect on facial and cranial base growth and are also odontogenic, but the effects have not been much studied. The growth-inhibiting effect of corticosteroids is explained partly by the reduced response of cartilage cells to insulin-like growth factor-1. Experimentally, the effect is also seen in the condylar cartilage, but clinical studies have not been published. In tooth eruption an accelerating effect has been noted in experimental animals. The role of androgenic hormones in mandibular growth stimulation is discussed.

□ *Glucocorticoids; growth hormone; human; sex steroids; thyroid hormones*

Sinikka Pirinen, P.O. Box 41, FIN-00014 University of Helsinki, Finland

Generally, it can be assumed that facial growth and general somatic growth are under the same endocrine control. This has been demonstrated in longitudinal studies comparing facial growth rates with those of somatic growth (1–3). Inside the craniofacial system, however, other growth mechanisms modify the timing and result of growth. Neural, lymphoid, and dental growth and the growth of upper airways follow their own course within the craniofacial bones, complicating the study of growth in the craniofacial area.

Endocrine regulation of growth has been studied by observing patients with endocrine disorders, analyzing samples from growing healthy children, and with animal experiments. Molecular biology techniques have opened new perspectives for this research and contributed significantly to the present understanding of growth regulation.

Growth hormone

Growth hormone (GH) is a peptide hormone secreted by the somatotrophic cells in the anterior lobe of the pituitary gland. Human growth hormone (hGH) differs from that of all other species. GH secretion is pulsatile, and the secretory bursts occur especially at early hours of sleep and throughout the night (4). The genes of the GH family are assigned to chromosome 17q23–q24 (5). The biologic actions of GH are mediated by its binding to a specific receptor located on the cells of target tissues. The GH receptor gene has also been characterized (6). GH has no specific target organ. It is an anabolic hor-

mone to which every organ system responds. However, it has a profound effect on cartilage growth. Many of its effects on bone and linear growth are believed to be mediated by insulin-like growth factors (IGFs) (7). The IGF-1 gene is located in chromosome 10 and is produced in the liver but also locally in tissues. Recently Ohlsson et al. (8) confirmed that GH is a primary and IGF-1 a secondary stimulatory effector in cartilage growth and maturation. GH is the main regulator of childhood and adolescent growth. Small children secrete less GH than tall ones (9). The GH pulse amplitude is increased in growth spurts. Plasma IGF-1 concentrations increase simultaneously.

Since 1985 hGH has been biosynthesized (rhGH) and, although very expensive, is available for the treatment of various growth disorders.

Reports on the dentition and facial structure of children who were GH-deficient (hypopituitary) started to appear in the dental literature in the 1930s and 1940s (10, 11). These reports, based on single cases, described delayed dental and facial development. Later, larger samples were analyzed, and delayed dental, facial, and cranial base development was demonstrated (12–14). Dental delay is always less pronounced than delay in height or bone age. In a group of 25 patients with growth hormone deficiency (GHD) we showed that the lag in dental development is progressive and varies between –1 SD and –6 SD (SD of dental age) (15). The dentition seems to be harmoniously delayed, so that all studied components of dental development (deciduous tooth resorption, permanent tooth formation, and eruptive movement) display the same degree

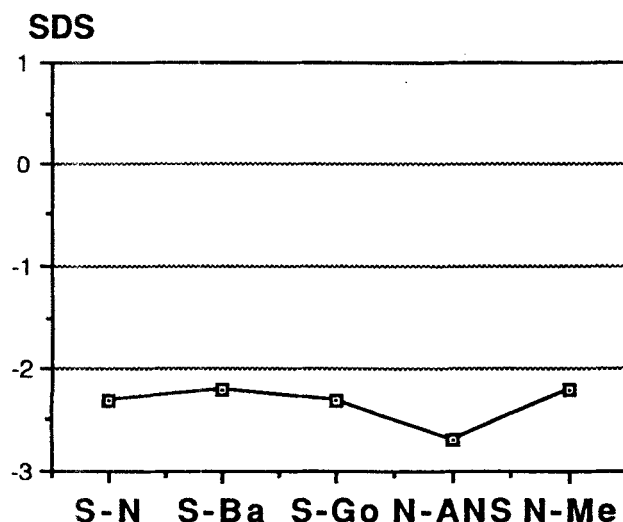


Fig. 1. Mean craniofacial pattern profile of 13 untreated patients with isolated or combined growth hormone deficiency. Mean age, 9.8 years. Age- and sex-specific means and standard deviations used in calculation of the standard deviation scores (SDS) by Bhatia & Leighton (60).

of retardation. GH starts to influence linear growth from about 9 months of age (16). Very little, therefore, is known about deciduous tooth eruption in GHD children. On the basis of our anamnestic findings, it appears to follow a normal or a slightly delayed timing. It is not yet clear whether treatment with GH influences dental development (15, 17, 18), but the results point towards a slight accelerating effect during the first years of treatment. The crown and root lengths are reduced in the hypopituitary dwarf mouse (19, 20). In the Snell mouse (19) all anterior pituitary hormones are lacking. The Lewis GHD dwarf rat displayed smaller populations of odontogenic cells and less mitoses in these cells (21). This experimental animal has very low GH production, whereas its other anterior pituitary hormone levels are normal.

Children with GHD are described as having a big skull and babyish face, which contrasts strongly with their intelligence, which is average for age. They are progressively shorter than their peers and often have a rich subcutaneous fat layer (16). Cephalometric studies have demonstrated small size of anterior and posterior cranial bases and small mandibular dimensions (13, 22). Our own analysis (23) of 21 patients with isolated or combined pituitary deficiency demonstrated small cranial base dimensions, small posterior facial height, and small posterior mandibular height, even when GHD patients were compared with healthy controls of the same height. In our group of 13 untreated patients (Fig. 1) with pituitary deficiency, the relatively smallest dimension in age-matched comparison was upper face height (N-ANS).

Excessive GH secretion leads to gigantism in the young and to acromegaly in adults. An intermediate form of patients, tall acromegals, is also recognized. GH hypersecretion is usually caused by a pituitary adenoma. We have described the craniofacial condition of two female patients (heights, 190 cm and 205 cm) with gigantism due to an excess of GH (23). Anterior facial heights were relatively the largest cephalometric dimensions (+6.8 and +3.3 SD), followed by posterior facial heights (+3.7 and +4.9 SD). Posterior cranial bases were long, but anterior cranial bases were of normal size. The faces of these female patients were broad, and the zygomatic arches pronounced. Cervical vertebrae were excessively high. Both had relatively normal dental occlusions.

Acromegaly is also rare but more common than gigantism. The serum IGF-1 levels of these patients are very high; the mean value of IGF-1 is reported to be tenfold higher than in normal adults (24). In this condition the hands, feet, and face grow characteristically at adult age. Mandibular growth is gradual and often noted by the dentist when crossbite has developed. The calvarium, hands, and feet grow by bone apposition. The skin and subcutaneous tissue thickens, reflecting the effect of GH in the connective tissue. The tongue grows, and general visceral overgrowth has been documented. Cartilaginous tissues enlarge, ribs thicken, and costochondral cartilage has been shown to be hypertrophic (25). Hypertrophic articular cartilage and the growth of chondrocytes in the articular cartilage may give rise to acromegalic arthropathy (26). It can be assumed that mandibular growth in acromegaly results from both appositional growth and hypertrophic changes in the condylar cartilage.

The treatment of different groups of patients with recombinant human growth hormone (rhGH) will in time give more evidence on dental and craniofacial growth in these patients, and orthodontists should therefore follow closely the results of these treatment studies. It will take some time before the final effect of GH treatment on craniofacial growth can be evaluated. A recent article, however, mentions acromegaloid features as one of the side effects of hGH treatment (27).

Thyroid hormones

Thyroid hormones (TH) lack a specific target organ; they affect every organ and system and every biologic process. Thyroid disorders are common, and their effects on craniofacial and dental structures are widely described in the literature. In prenatal hypothyroidism linear growth at birth is normal, but the development of bone and teeth is retarded (28). Later, enamel defects are seen in prenatally developed teeth. After birth hypothyroidism (HT) disturbs normal growth, resulting in growth retardation or complete cessation of growth in athyroidism. The development and growth of the central

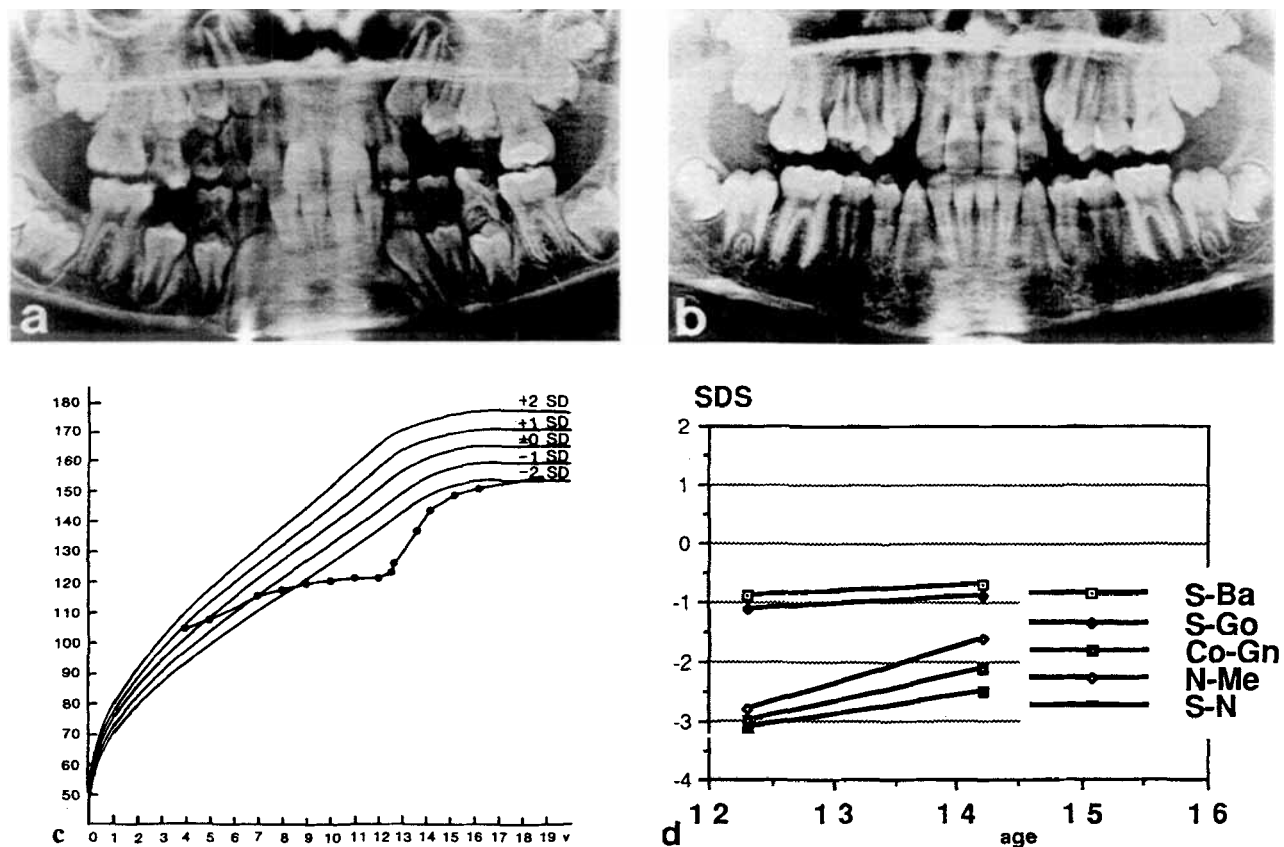


Fig. 2. Panoramic radiograph of a girl with hypothyroidism at diagnosis (12.2 years) (a) and after 14 months of replacement therapy (13.3 years) (b). Dental development is delayed; dental age based on tooth formation (61, 62) is 10.2 years (-2.2 SD) and 11.3 years (-2.0 SD). Eruption is more delayed than formation: permanent canines and premolars show two-thirds root development without corresponding eruptive movement or resorption of deciduous teeth. Note rapid recovery with therapy. 2c) Growth curve of the patient. 2d) Craniofacial dimensions at start and after 2 years of replacement therapy.

nervous system is seriously damaged. Growth of the cranium is retarded, and characteristic brachycephalic facies develops (29). Dental age is less delayed than bone age (13, 14). Rapid recovery of dental delay has been seen when treatment with thyroid hormone has been instituted (Fig. 2a,b) (30).

The presence of thyroid hormones is important in the synthesis of IGF-1. It is likely that thyroid hormone enhances cartilage growth through IGF-1 and by directly accelerating the differentiation of chondrocytes (31). The dental and craniofacial retardation found in hypothyroid conditions of longer duration differs somewhat from that seen in isolated lack of GH. The main difference is in the cranial vault, which shows a growth retardation in HT. Reduced facial height is typical in children with untreated HT of long duration (Fig. 2d). While retardation of tooth formation is of the same order in the two endocrinopathies, arrested tooth eruption is seen in HT. It has been suggested that this effect might be related to the epidermal growth factor (EGF) or EGF receptors (32, 33).

Some old textbooks show pictures of children with hyperthyroidism and demonstrate very early tooth eruption (34), whereas some reports (14) indicate delayed dental development.

Sex steroids

A slight increase in growth rate is seen at the age of 6–8 years in most children (the mid-childhood growth spurt). This is thought to be due to increased GH and IGF-1 production, which is stimulated by adrenal androgens. At puberty an increase in GH production is seen in boys and girls. Several lines of evidence indicate that this increase is sex-steroid-dependent. The sex steroids here are adrenal and ovarian androgens and ovarian and testicular estrogens. Plasma concentrations of IGF-1 show a similar increase (35). In addition to the increase in GH and IGF-1 levels, sex steroids display a small, direct effect on linear growth. This effect is especially seen as vertebral growth (36).

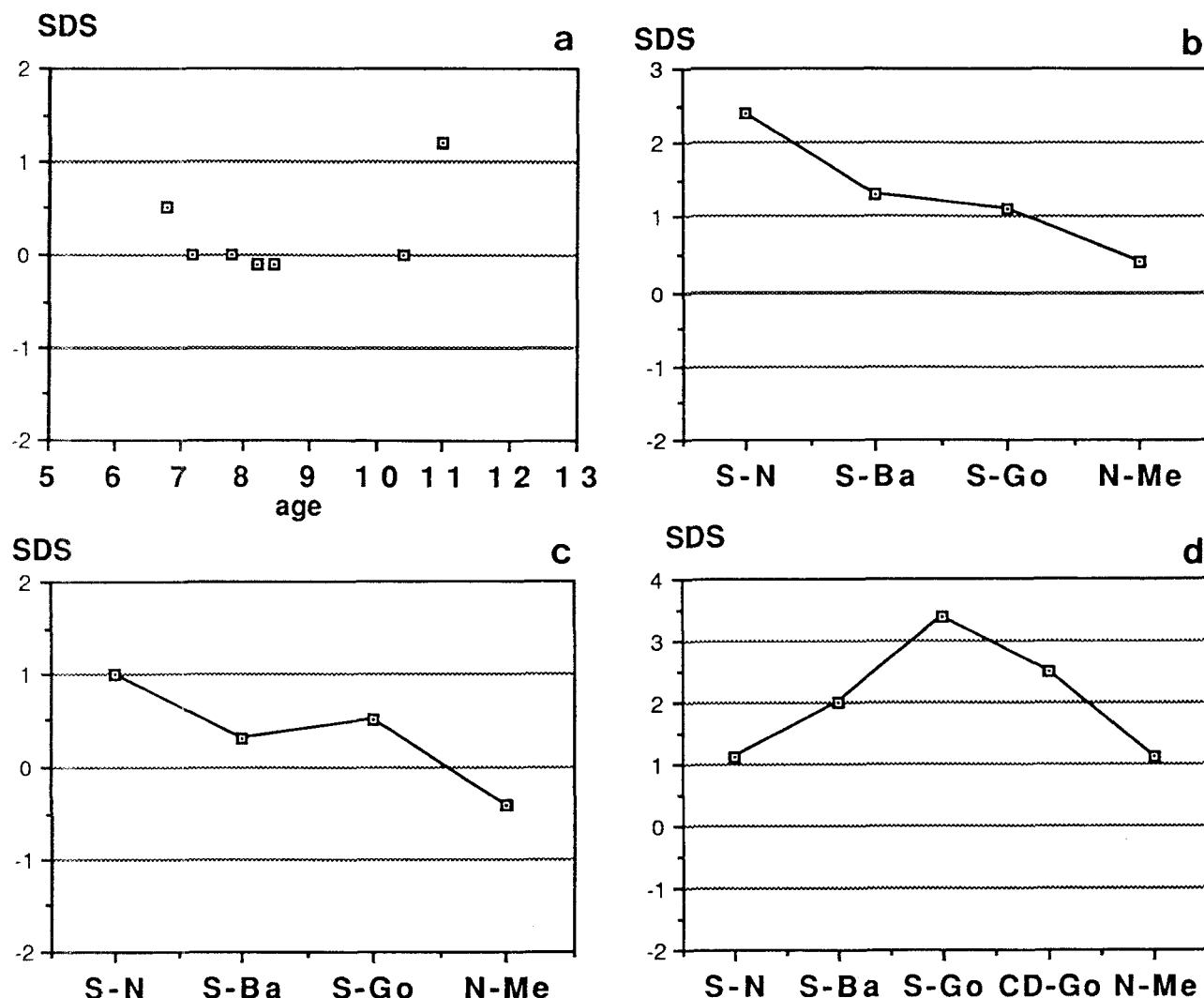


Fig. 3a) Standard deviation scores (SDS) of dental ages of seven patients with precocious puberty (six girls, one boy, aged 7.2 years). 3b) Average craniofacial pattern profile of the four younger girls (bone ages 11, 11.5, 12, and 11 years). 3c) The two older girls (bone age, 16 years) and 3d) the boy (bone age, 13 years). Cranial base and facial height dimensions are advanced in the younger girls and in the young boy and also in the boy's mandibular size, suggesting androgen sensitivity in mandibular growth regulation.

Again, valuable information on the role of sex steroids in dental and craniofacial development has been obtained by studying children with disorders of puberty. In normal, but extremely early or late-maturing children, dental development showed a slight deviation to early or late development, correspondingly (13). On the basis of these observations, Garn et al. (12) noted some sex steroid dependence in late-forming teeth. This finding has not been uniformly confirmed by others, but it reminds us that the different components of the dentition and their development should perhaps be analyzed separately to fully understand the regulation of dental development.

In true sexual precocities a slight advancement of dental age is reported (14). Too far-reaching conclusions

should not be drawn on these endocrinopathies as they are seldom of long duration (Fig. 3a).

Advanced facial and cranial base growth has been noted in cases of sexual precocity (Fig. 3b-d). This advancement has, however, been less pronounced than advancement of bone age (13). Correspondingly, delayed facial growth has been demonstrated in delayed puberty and hypogonadism.

In congenital adrenal hyperplasia, increased adrenal androgen secretion causes advanced craniofacial growth. Advanced dental development is also seen (12-14, 37). However, the dentition is always much less advanced than bone age.

Animal experiments with anabolic steroids have demonstrated a clear effect on craniofacial growth,

mainly as overgrowth and rotation of the maxilla (38, 39). Barrett & Harris (38) present the assumption that misuse of anabolic steroids by adolescents and athletes may produce changes also in their craniofacial structures.

Thus, orthodontists should be alert also in following up research related to anabolic steroids and their therapeutic use and abuse.

Corticosteroids

Hyperglucocorticoidism leads to short stature and delayed bone maturation but increases relative weight (40). Very small amounts of cortisol given as medication can decrease growth rate. The mechanism is explained on the basis of lowered GH secretion, and the response of cartilage cells to IGF-1 is also lowered. Moreover, skeletal IGF-1 synthesis is decreased by cortisol, which has an inhibitory effect on bone collagen synthesis (41). In animal and tissue culture experiments mandibular condylar cartilage seems to respond to cortisol in a manner similar to the growth plates of long bones (42–44). Tissues in which cells continuously and rapidly reform (intestinal mucosa, testes, spleen, bone marrow) are more resistant to the 'glucocorticoid retarding effect' (45).

In the process of tooth eruption, however, cortisone has a special effect (46). Eruption rate is accelerated, but the mechanism behind this phenomenon is still uncharacterized. When studying the rat incisor, Teng et al. (47) noted accelerated tooth eruption after cortisone administration only if the fundus of the erupting tooth was left intact.

There seem to be no controlled clinical studies of facial growth or tooth eruption rate in lack or excess of cortisone. There could be an effect, for example, when children with juvenile rheumatoid arthritis are treated with systemic cortisol, when local treatment with cortisone for severe eczema is used, or in connection with inhalation medication for asthma (48; Westphal, personal communication, 1994). The adjustment of doses to an alternate-day regimen has reduced the growth-inhibiting effect of pharmacologic cortisone treatment (49).

Discussion

All in all, facial growth is very much under general growth control, and GH seems to be an important factor also in facial growth.

The determinants of dental development are less clear and hardly to be found by studying classical hormones. GH and TH no doubt, have an obvious effect on dental development. The lack of these two hormones seems to affect the timing of development of the dentition, whereas it is unclear whether an excess of GH or TH

has an effect on dental development. The effect may be related to the general mitogenic effects of GH and IGF-1. TH seems to have some role of its own in the eruptive movement of teeth. When experimental work is evaluated, it must be kept in mind that the effects of TH on GH gene expression and GH synthesis vary significantly among mammalian species (50).

Adrenal androgens seem to be odontogenic, but the phenomenon and the mechanism behind it should be better studied.

Clearly, mandibular growth is controlled by GH. Several lines of evidence suggest that mandibular growth is also sensitive to direct androgen stimulation, in addition to the fact that sex steroids exert their effect by increasing GH secretion. Studies clearly demonstrate a pronounced mid-childhood spurt in mandibular growth (1, 3, 51–53), whereas growth in height is only slightly accelerated (54). The mid-childhood growth spurt is produced by adrenal androgens in boys and girls. At puberty mandibular growth peaks a little later than height. A considerable part of the height gain at puberty results from the rapid growth of the vertebrae (55). The peak height velocity (PHV) for sitting height comes later than the PHV for leg length. Vertebral growth is said to be under direct androgen influence, and the later timing of maximal mandibular growth could also reflect the same dependency. Vertebral growth continues slowly to the third decade of life. The timing suggests that androgenic hormones may be of importance in maintaining this growth (55). The mandible also grows slowly during adult life (56). The third line of evidence suggesting androgen sensitivity in mandibular growth control is the obvious sexual dimorphism in human facial dimensions. The male mandibular ramus is on the average 14% longer than the female ramus, whereas other facial dimensions have approximately an 8% sex difference (57).

Orthodontists should carefully follow the new developments in pharmacologic growth stimulation, treatments with classical hormones, and local treatment with GH, IGF-1, and other growth factors (58, 59). When the problem of the local application of hGH and growth factors is successfully solved, new possibilities for the treatment of craniofacial growth problems will open.

Acknowledgements.—The author presents her warm thanks to Professor Jaakko Perheentupa and Dr. Hanna-Liisa Lenko, Children's Hospital, University of Helsinki, Division of Pediatric Endocrinology, for cooperation and guidance.

References

1. Nanda RS. The rates of growth of several facial components measured from serial cephalometric roentgenograms. *Am J Orthod* 1955;41:653–73.
2. Björk A, Helm S. Prediction of age of maximum body height. *Angle Orthod* 1967;37:134–43.
3. Baughan B, Demirjian A, Levesque GY, La Palme-Chaput L.

- The pattern of facial growth before and during puberty as shown by French-Canadian girls. *Ann Hum Biol* 1979;6:59-76.
4. Finkelstein J, Rofwarg H, Boyar A. Age-related changes in the 24-hour spontaneous secretion of growth hormone. *J Clin Endocrinol Metab* 1972;26:1173-9.
 5. Chen EY, Liao Y-C, Smith DH, Barrera-Saldana HA, Gelinas RE, Seeburg PH. The human growth hormone locus: nucleotide sequence, biology, and evolution. *Genomics* 1989;4:479-97.
 6. Leung D, Spencer S, Cachianes GR. Growth hormone receptor and serum binding protein: purification, cloning and expression. *Nature* 1987;330:537-44.
 7. Daughaday WH, Rotwein P. Insulin like growth factors -1 and -2. Peptide, messenger, ribonucleic acid and gene structures, serum and tissue concentrations. *Endocrine Rev* 1989;10:68-91.
 8. Ohlsson C, Isaksson O, Lindahl A. Clonal analysis of rat tibial growth plate chondrocytes in suspension culture—differential effects of growth hormone and insulin-like growth factor I. *Growth Regul* 1994;4:1-7.
 9. Albertsson-Wikland K, Rosberg S. Analyses of 24 hour growth hormone profiles in children: relation to growth. *J Endocrinol Metab* 1988;67:493-500.
 10. Schour I, Brodie A, King E. The hypophysis and teeth: a case report. *Angle Orthod* 1934;4:285-304.
 11. Cohen MM, Wagner R. Dental development in pituitary dwarfism. *J Dent Res* 1948;27:445-51.
 12. Garn SM, Lewis AB, Blizzard RM. Endocrine factors in dental development. *J Dent Res* 1965;44:243-58.
 13. Spiegel R, Sather H, Hayles A. Cephalometric study of children with various endocrine diseases. *Am J Orthod* 1971;59:362-75.
 14. Keller EE, Sather AH, Hayles AB. Dental and skeletal development in various endocrine and metabolic diseases. *J Am Dent Assoc* 1970;81:415-9.
 15. Myllärniemi S, Lenko H-L, Perheentupa J. Dental maturity in hypopituitarism and dental response to substitution treatment. *Scand J Dent Res* 1978;86:307-12.
 16. Karlberg J, Albertsson-Wikland K. Infancy growth pattern related to growth hormone deficiency. *Acta Paediatr Scand* 1988; 77:385-91.
 17. Sarnat H, Kaplan I, Pertzalan, Laron Z. Comparison of dental findings in patients with isolated growth hormone deficiency treated with human growth hormone (hGH) and in untreated patients with Laron type dwarfism. *Oral Surg Oral Med Oral Pathol* 1988;66:581-6.
 18. Ito RK, Vig KW, Garn SM, Hopwood N, Loos PJ, Spalding PM, et al. The influence of growth hormone (rhGH) therapy on tooth formation in idiopathic short statured children. *Am J Orthod Dentofac Orthop* 1993;103:358-64.
 19. Snell GD. Dwarf. A new mendelian recessive character of the house mouse. *Proc Natl Acad Sci USA* 1929;15:733-4.
 20. Tracey C, Roberts GJ. A radiographic study of dental development in the hypopituitary dwarf mouse. *Arch Oral Biol* 1985;30:805-11.
 21. Young WG, Zhang CZ, Li H, Osborne P, Waters MJ. The influence of growth hormone on cell proliferation in odontogenic epithelia by bromodeoxyuridine immunohistochemistry and morphometry in the Lewis dwarf rat. *J Dent Res* 1992;71:1807-11.
 22. Poole AE, Greene IM, Buchang PH. The effect of growth hormone therapy on longitudinal growth of the oral facial structures in children. In: Factors and mechanisms influencing bone growth. New York: Alan R Liss, 1982:499-516.
 23. Pirinen S, Majurin A, Lenko H-L, Koski K. Craniofacial features in lack and excess of growth hormone. *J Craniofac Gen Devel Biol* 1994;14:144-52.
 24. Clemmons D, van Wyk J, Ridgeway E. Evaluation of acromegaly by radioimmunoassay of somatomedin-C. *N Engl J Med* 1979;301: 1138-42.
 25. Merimee TJ, Grant MB. In: Becker K, editor. Principles and practice of endocrinology and metabolism. Philadelphia: JB Lippincott, 1990:125-33.
 26. Podgorski M, Robinson B, Weusberger A. Articular manifestations of acromegaly. *Aust NZ J Med* 1988;18:28-32.
 27. Neely EK, Rosenfeldt RG. Use and abuse of growth hormone. *Annu Rev Med* 1994;45:407-20.
 28. Andersen HJ. Studies in hypothyroidism in children [thesis]. Copenhagen: University of Copenhagen, 1961.
 29. Caffey J. Pediatric X-ray diagnosis. 3rd ed. Chicago: Year Book Medical Publishers, Inc, 1957.
 30. Reuland-Bosma W, Dibbets JM. Mandibular and dental development subsequent to thyroid therapy in a boy with Down syndrome: report of a case. *J Dent Child* 1991;58:64-8.
 31. Burch WM, van Wyk JJ. Triiodothyronine stimulates cartilage growth and maturation by different mechanisms. *Am J Physiol* 1987;252:176-82.
 32. Cohen S. Isolation of a mouse submaxillary gland protein accelerating incisor eruption and eyelid opening in newborn animals. *J Biol Chem* 1962;237:1555-62.
 33. Alm J, Lakshamanan J, Hoath S, Fisher DA. Neonatal hyperthyroidism alters hepatic epidermal growth factor receptor ontogeny in mice. *Pediatr Res* 1988;23:557-60.
 34. Salzmann JA. Principles of orthodontics. 2nd ed. Philadelphia: JB Lippincott, 1943:343.
 35. Cara JF. Growth hormone in adolescence, normal and abnormal. *Endocr Metab Clin N Am* 1993;22:533-52.
 36. Cara JF, Kreiter ML, Rosenfield RL. Height prognosis of children with true precocious puberty and growth hormone deficiency: effect of combination therapy with gonadotrophin releasing hormone agonist and growth hormone. *J Pediatr* 1992;120:709-14.
 37. Wagner R, Cohen MM, Hunt EE. Dental development in idiopathic sexual precocity, congenital adrenocortical hyperplasia and adrenogenic virilism. *J Pediatr* 1963;63:566-75.
 38. Barrett RL, Harris EF. Anabolic steroids and craniofacial growth in the rat. *Angle Orthod* 1993;63:289-98.
 39. Noda K, Chang H-P, Takahashi R, Kinoshita Z, Kawamoto T. Effects of the anabolic steroid nandrolone phenylpropionate on craniofacial growth in rats. *J Morphol* 1994;220:25-33.
 40. MacArthur RG, Cloutier MD, Hayles AB. Cushing's disease in children. *Mayo Clin Proc* 1972;47:318-26.
 41. McCarthy T, Centrella M, Canalis E. Cortisol inhibits the synthesis of insulin-like growth factor-1 in skeletal cell. *Endocrinology* 1990;126:1569-75.
 42. Hsien C-K, Johannessen L. Skeletal changes in cortisone treated male rats. *J Dent Res* 1970;49:34-41.
 43. Davidovitch Z. Radiographic and autoradiographic study on the effects of cortisone on bone growth in young albino rats. *Arch Oral Biol* 1971;16:879-914.
 44. Maor G, Silberman M. Studies of hormonal regulation of the growth of the craniofacial skeleton. IV. Specific binding sites for glucocorticoids in condylar cartilage and their involvement in the biological effects of glucocorticoids on cartilage cell growth. *J Craniofac Gen Devel Biol* 1986;6:189-202.
 45. Ballard PL. Glucocorticoids and differentiation. In: Baxter JD, Rousseau GC, editors. Glucocorticoid hormone action. New York: Springer-Verlag, 1979:493.
 46. Domm LV, Wellband WA. Effect of adrenalectomy and cortisone on eruption rate of incisors in young female albino rats. *Proc Soc Exp Biol Med* 1960;104:582-4.
 47. Teng C-M, Sobkowski J, Johnston L Jr. The effect of cortisone on the eruption rate of root resected incisors in the rat. *Am J Orthod Dentofac Orthop* 1989;95:67-71.
 48. Turpeinen M, Salo OP, Leisti S. Effect of percutaneous absorption on adrenocortical responsiveness in infants with severe skin disease. *Br J Dermatol* 1986;115:475-84.
 49. Soyka LF. Treatment of the nephrotic syndrome in childhood. *Am J Dis Child* 1967;113:693-701.
 50. Brent GA, Harney JW, Moore DD, Larsen PR. Multihormonal regulation of the human, rat, and bovine growth hormone promoters: differential effect of 3',5'-cyclic adenosine monophosphate, thyroid hormone and glucocorticoids. *Mol Endocrinol* 1988;2:792.

51. Woodside DG, Reed RT, Doucet RT, Thompson GW. Some effects of activator treatment on the growth rate of the mandible and the position of the midface. Transactions of the 3rd Orthodontics Congress. St Louis: CV Mosby Co., 1975.
52. Ekström C. Facial growth rate and its relation to somatic maturation in healthy children. *Swed Dent J* 1982 Suppl 11.
53. Krieg WL. Early facial growth accelerations. A longitudinal study. *Angle Orthod* 1987;57:50–62.
54. Meredith HV. An addendum on presence and absence of a mid-childhood spurt in somatic dimensions. *Ann Hum Biol* 1981; 8:473–76.
55. Tanner JM. The human growth curve. In: Harrison GA, Weiner JS, Tanner JM, Barnicot NA, editors. *Human biology*. 2nd ed. Oxford: Oxford University Press, 1977:299–319.
56. Behrents RG. The consequences of adult craniofacial growth. In: Carlson DS, editor. *Orthodontics in an aging society*. Ann Arbor (MI): Center for Human Growth and Development, University of Michigan, 1989:53–99. Monograph No. 22, Craniofacial Growth Series.
57. Hunter WS, Garn SM. Disproportionate sexual dimorphism in the human face. *Am J Phys Anthr* 1972;36:133–8.
58. Isgaard J, Nilsson A, Lindahl A, Jansson J-O, Isaksson O. Effects of local administration of GH and IGF-1 on longitudinal bone growth in rats. *Am J Physiol* 250 (Endocrinol Metab 13): E367–72.
59. Nilsson A, Isgaard J, Lindahl A, Peterson L, Isaksson O. Effects of unilateral arterial infusion of GH and IGF-1 on tibial longitudinal bone growth in hypophysectomized rats. *Calcif Tissue Int* 1987;40:91–6.
60. Bhatia SN, Leighton BC. *A manual of facial growth. A computer analysis of longitudinal cephalometric growth data*. Oxford: Oxford University Press, 1993.
61. Demirjian A, Goldstein H. New systems for dental maturity based on seven and four teeth. *Ann Hum Biol* 1976;3:411–21.
62. Kataja M, Nyström M, Aine L. Dental maturity standards in southern Finland. *Proc Finn Dent Soc* 1989;85:187–97.