

## Correspondence and Short Communications

*Comments on published articles, short communications of a preliminary nature, case reports, technical notes and the like are accepted under this heading. The articles should be short and concise and contain a minimum of figures, tables and references.*

### UNBOUND, BIOLOGICALLY ACTIVE OESTRADIOL, SEX HORMONE-BINDING GLOBULIN, EPIDERMAL GROWTH FACTOR AND ELECTROLYTES IN BREAST CYST FLUID

Several studies have shown that women with gross cystic breast disease (cysts of more than 3 mm in diameter), a common condition affecting about 7% of women in the western hemisphere, may be at increased risk of developing breast cancer (1, 2). Since both oestradiol ( $E_2$ ) and epidermal growth factor (EGF) may play a role in the development of breast cancer (3, 4), we thought it pertinent to measure the concentrations of unbound, biologically active  $E_2$  and EGF in breast cyst fluid. In addition, we studied the relationship between intracystic concentrations of  $E_2$  and sex hormone-binding globulin (SHBG) as this has not been reported in the literature.

**Material and Methods.** Breast cyst fluid was obtained by needle aspiration of 25 breast cysts from 25 women (aged 30 to 60 years with a median age of 45 years) who attended the Breast Clinic at St. Mary's Hospital, London. The samples were centrifuged and the supernatant stored at  $-20^\circ\text{C}$  until analysis.

Anti- $E_2$  antiserum raised against oestradiol-6(0-carboxymethyl)-oxime bovine serum albumin in goats was a gift from Dr. G. Knaggs. Anti-human EGF antiserum and pure urogastone (EGF isolated from human urine) was purchased from Amersham International plc, Amersham, Bucks., UK. The SHBG immunoradiometric kit was obtained from Farnos Diagnostica Ltd., Organon Teknika Ltd., Cambridge, UK. The Dianorm equilibrium dialysis system was obtained from MSE Scientific Instruments, Crawley, UK.

Total  $E_2$  was estimated with a method previously described (5). The sensitivity of the assay was 18 pmol/l and the intra-assay and interassay coefficients of variation were 6.5% and 10.7% respectively.

The percentage of unbound  $E_2$  in breast cyst fluid was measured using a dianorm equilibrium dialysis machine. This methodology has been described earlier (6). The principle of the technique is to dialyse breast cyst fluid against (2, 4, 6, 7- $^3\text{H}$ ) $E_2$  (approximately 10 000 cpm) until equilibrium is reached. The unbound  $E_2$  concentration was calculated by multiplying the percentage of unbound  $E_2$  by the total  $E_2$  concentration. The intra-assay and interassay coefficients of variation were 3.7% and 12.9% respectively.

SHBG was measured by an immunoradiometric method (Farnos Diagnostica kit). Breast cyst fluid was assayed in 2 dilutions, 1 in 5 and 1 in 10. The sensitivity of the assay was 0.5 nmol/l. The intra-assay and interassay coefficients of variation were 2.1% and 6.5% respectively.

EGF was measured by a double antibody radioimmunoassay. Sample/standard (0.1 ml) was incubated with 0.1 ml of label (about 8 000 cpm), 0.1 ml of antiserum (working dilution of 1 in 30 000) and 0.2 ml of assay buffer (0.05 mol/l phosphate buffer, pH 7.4, containing 0.2% bovine serum albumin) at  $4^\circ\text{C}$  for 3 days with a further day's incubation following the addition of donkey anti-rabbit second antibody.

The sensitivity of the assay was 0.04 ng/ml and the intra-assay and interassay coefficients of variation were less than 10%.

Sodium and potassium concentrations were measured using an indirect ion selective electrode (Beckman Electrolyte 2 Analyser).

The data were not normally distributed. Non-parametric statistics were, therefore, used. Correlations were assessed using Spearman's rank correlation method ( $r_s$  = Spearman's rank correlation coefficient). Results were considered to be statistically significant when  $p < 0.05$ .

**Results.** The concentration ranges and median concentrations of  $E_2$  (total and unbound), SHBG, EGF, sodium and potassium are shown in the Table.

No correlation was found between the intracystic concentration of total  $E_2$  and SHBG. A negative correlation ( $r_s = -0.431$ ,  $p < 0.05$ ) was obtained between the intracystic concentrations of unbound  $E_2$  and SHBG (Figure). No correlations were found between intracystic concentrations of EGF and total or unbound  $E_2$ .

**Discussion.** The exact mechanism by which SHBG, a protein which is synthesised in the liver, enters breast cyst fluid from plasma remains an enigma. Entry into cyst fluid, however, appears to be better in cysts with high intracystic sodium to potassium ratios. A positive correlation was obtained between SHBG

**Table**

*Concentration ranges and median values of analytes measured (n = 25)*

Analyte	Range	Median
Total $E_2$	50–1 000 pmol/l	103 pmol/l
Unbound $E_2$	4.3–119 pmol/l	9.7 pmol/l
SHBG	1.3–143 nmol/l	3.5 nmol/l
EGF	1.8–570 ng/ml	178 ng/ml
Sodium	27–145 mmol/l	128 mmol/l
Potassium	4–124 mmol/l	19 mmol/l

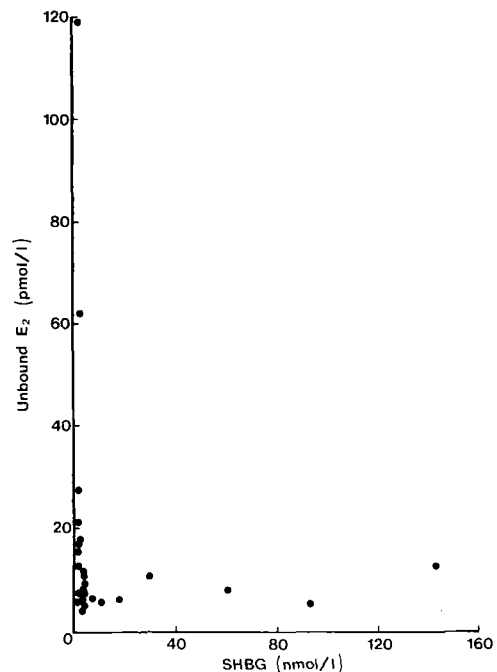


Figure. Concentrations of unbound  $E_2$  versus SHBG ( $r_s = -0.431$ ,  $p < 0.05$ ,  $n = 25$ ).

concentrations and sodium to potassium ratios in breast cyst fluid ( $r_s = 0.453$ ,  $p < 0.02$ ). Intracystic sodium to potassium ratios may reflect the nature of the epithelial lining of breast cysts, ratios of less than three may indicate the presence of apocrine epithelial lining while ratios of greater than three may indicate the presence of attenuated epithelial lining (7).

Concentrations of testosterone and dihydrotestosterone have been shown to be higher in breast cyst fluid than total oestradiol (8). In cysts with low intracystic sodium to potassium ratios, where SHBG concentrations are lower, the presence of high concentrations of testosterone and dihydrotestosterone, both of which bind more strongly to SHBG, may be the explanation for the higher proportion of unbound  $E_2$  present in these cysts.

$E_2$  in breast cyst fluid may be derived by metabolism of dehydroepiandrosterone sulphate by the cyst wall (9). EGF was present in high concentrations in breast cyst fluid, supporting the findings of Jaspar & Franchimont (10). Concentrations of EGF are almost undetectable in plasma (11). It seems likely that EGF in breast cyst fluid may arise from local synthesis and secretion of EGF by mammary epithelial cells rather than the active transport of EGF from plasma into cyst fluid. Wide-ranging concentrations of total and unbound  $E_2$  and EGF were found in breast cyst fluid. The significance of this finding is unknown.

In view of the possible role which both unbound, biologically active  $E_2$  and EGF may have in carcinogenesis, we postulate that women who have high intracystic concentrations of both unbound  $E_2$  and EGF may be at higher risk of breast cancer. Longitudinal studies on women with gross cystic breast disease assessing the predictive value of intracystic concentrations of  $E_2$  and EGF should perhaps be carried out.

*Key words:* Breast, cyst fluid, oestradiol, sex hormone-binding globulin, epidermal growth factor.

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## REFERENCES

1. Azzopardi JB. Problems in breast pathology. Philadelphia: WB Saunders, 1979: 92–112.
2. Haagensen CD, Bodian C, Haagensen DE. Breast carcinoma: Risk and detection. Philadelphia: WB Saunders, 1981.
3. James VHT, Reed MJ. Steroid hormones and human cancer. In: Iacobelli S, King RJB, Lindner HR, Lippman ME, eds. Hormones and cancer. New York: Raven Press, 1980: 471–87.
4. Stoscheck CM, King LE. Role of epidermal growth factor in carcinogenesis. *Cancer Res* 1986; 46: 1030–7.
5. Braunsberg H, Reed MJ, Short F, Dias VO, Baxendale PM. Changes in plasma concentrations of oestrogens and progesterone in women during anaesthesia and gynaecological operations. *J Steroid Biochem* 1981; 14: 749–55.
6. James VHT, Reed MJ, Folkard EJ. Studies of oestrogen metabolism in postmenopausal women with cancer. *J Steroid Biochem* 1981; 15: 235–46.
7. Dixon JM, Miller WR, Scott WN, Forrest APM. The morphological basis of human breast cyst populations. *Br J Surg* 1983; 70: 604–6.
8. Bradlow HL, Rosenfeld RS, Fleisher M, O'Connor J, Schwartz MK. Steroid hormone accumulation in human breast cyst fluid. *Cancer Res* 1981; 41: 105–7.
9. Bradlow HL, Schwartz MK, Fleisher M, et al. Hormone levels in human breast cyst fluid. In: Angeli A, Bradlow HL, Dogliotti L, eds. Endocrinology of cystic breast disease. New York: Raven Press, 1983: 59–75.
10. Jaspar JM, Franchimont P. Radioimmunoassay of human epidermal growth factor in human breast cyst fluid. *Eur J Cancer Clin Oncol* 1985; 21: 1343–8.
11. Oka Y, Orth DN. Human plasma epidermal growth factor/ $\beta$ -urogastrone is associated with blood platelets. *J Clin Invest* 1983; 72: 249–59.