

ORIGINAL ARTICLE

## Effect of dentine moisture conditions on interfacial adaptation of a calcium silicate-based sealer compared with an epoxy resin-based sealer: an SEM/EDX analysis

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### ABSTRACT

**Introduction:** To evaluate the effect of dentine moisture conditions on interfacial adaptation of a calcium silicate-based sealer (Bio-C Sealer) compared with that of an epoxy resin-based sealer (AH Plus Jet) at different root levels.

**Methods:** Sixty extracted single-rooted teeth were prepared using ProTaper Universal system up to size F3 and randomly allocated to two groups ( $n = 30$ ) according to the type of sealer. Each group was then further subdivided into three subgroups ( $n = 10$  each) according to dentine moisture condition: dry (ethanol as final irrigant), normal moist (blotted dry), and wet (canals left flooded). Root sections (2 mm in thickness) were obtained at 3 mm (apical) and 8 mm (coronal). Interfacial adaptation was assessed using scanning electron microscopy (SEM) analysis, while elemental composition was analyzed utilizing energy-dispersive X-ray spectroscopy analysis (EDX). Data were analyzed using two-way analysis of variance and Tukey's post hoc test ( $p \leq 0.05$ ).

**Results:** Dentine moisture significantly affected interfacial adaptation. AH Plus Jet demonstrated superior adaptation under dry conditions, while Bio-C Sealer demonstrated optimal adaptation under moist conditions. EDX analysis revealed significantly higher levels of calcium and phosphorus in the Bio-C Sealer compared with the AH Plus group ( $p \leq 0.019$ ;  $p \leq 0.036$ ). Within the Bio-C Sealer group, calcium levels were significantly lower coronally ( $p = 0.016$ ) than apically. Silicon and carbon were higher in AH Plus ( $p < 0.01$ ), while oxygen was higher in Bio-C ( $p < 0.001$ ).

**Conclusion:** Dentine moisture conditions significantly influence the performance of sealer. AH Plus performs optimally under dry conditions, while Bio-C Sealer demonstrates superior performance in moist environments.

### ARTICLE HISTORY

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### KEYWORDS

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adaptation; sealer

### Introduction

Based on the classification of endodontic materials, epoxy resin-based sealers, particularly AH Plus Jet (Dentsply Sirona; Ballaigues; Switzerland), have been considered as the gold standard due to their reliable clinical performance and favorable physicochemical properties [1]. Despite these advantages, this sealer exhibits intrinsic biological and interfacial limitations that warrant critical evaluation [2, 3].

From an interfacial perspective, the hydrophobic nature of the epoxy resin-based sealers justifies a well-controlled dry environment to accomplish optimal marginal adaptation [4, 5]. Residual dentine moisture may interfere with the polymerization, minimize penetration of sealers into the dentinal tubules, and promote interfacial gap formation [6]. Such limitations have led to the development of bioactive alternatives, especially calcium silicate-based sealers, which are formulated to function optimally in the presence of moisture. Calcium silicate-based (bioceramic) sealers interact with the dentinal tissues through

hydration reactions that benefit from environmental moisture to produce calcium silicate hydrate and calcium hydroxide. These reaction products enhance the formation of apatite-like precipitates at the dentine–sealer interface, thus improving the biological seal and interfacial integrity [7].

Among calcium silicate-based sealers, Bio-C Sealer (Angelus; Londrina, PR, Brazil) has gained significant interest due to its premixed formulation and bioactive characteristics [8]. Previous studies have recorded its ability to penetrate into the dentinal tubules and promote mineralized interfacial layer formation, potentially enhancing its ability to seal the root canal system [9–12]. Despite these favorable properties, the performance of calcium silicate-based sealers is greatly dependent on the moisture condition of the root canal. Variations in dentine moisture, ranging from dry to moist or excessively wet, may affect hydration kinetics, ion release, and subsequent biomineralization [13, 14]. While some researchers have investigated the sealing ability and bonding strength of bio-ceramic sealers

[14, 15] and their biocompatibility [16], limited data are available regarding the combined morphological and elemental characteristics of the dentine–sealer interface under different dentine moisture conditions.

Additionally, although the bioactivity of calcium silicate-based sealers has been widely investigated [14–17], limited evidence is available with regard to how controlled dentine moisture may affect their interfacial behavior in comparison with a conventional epoxy resin-based sealer. Given that clinical canal drying protocols vary and that the bio-ceramic materials differ fundamentally from resin-based sealers in their hydration interaction with dentine moisture, understanding these interactions is essential.

Thus, this study aimed to assess the influence of dentine moisture (dry, normal moist, and wet) on the interfacial adaptation and elemental composition of a calcium silicate-based sealer (Bio-C Sealer) in comparison to an epoxy resin-based sealer (AH Plus Jet) at the apical and coronal root canal levels, using combined scanning electron microscopy/energy-dispersive X-ray spectroscopy (SEM/EDX) analysis. The tested null hypotheses were:

1. Dentine moisture condition would not significantly influence the interfacial adaptation or elemental composition of the investigated sealers.
2. No significant differences would exist between the two root canal sealers in relation to interfacial adaptation and elemental composition, regardless of the dentine moisture condition or root canal region.

## Materials and methods

### Sample size determination

Sample size was calculated using G\*Power software (version 3.1.9.7; Heinrich-Heine-Universität, Düsseldorf, Germany) based on a two-way analysis of variance (ANOVA) design. Assuming a medium effect size ( $f = 0.30$ ), a statistical power of 0.80, and a significance level of  $\alpha = 0.05$ . The minimum required sample size was 60 samples.

Sixty single-rooted human mandibular premolars with fully developed apices were collected from patients aged 18–35 years following extraction for orthodontic reasons. The teeth were cleaned of debris and stored in 0.1% thymol solution at 4°C for a maximum of 3 months to maintain dentine hydration [5].

### Root canal preparation

Teeth were sectioned at the level of the cemento-enamel junction using a low-speed diamond saw (Isomet 4000; Buehler, USA) under water cooling to obtain a standardized root lengths of  $14 \pm 1$  mm. Working length was determined by inserting a #10 K-file until it became visible at the apical foramen, and then 1 mm was subtracted from the measured root length. All root canals were instrumented using the ProTaper Universal rotary system up to size F3 (30/.09) [5]. Irrigation protocol was performed by using 3 mL of 5% sodium hypochlorite (NaOCl) after

each file with a 30-gauge side-vented needle, followed by 5 mL of 17% Ethylenediaminetetraacetic acid (EDTA) that was applied for 3 minutes to remove a smear layer, and then a final flush with distilled water was used [9].

### Dentinal moisture protocols

The manufacturer-reported classification and the chemical composition of the investigated sealers are detailed in Table 1.

The specimens were randomly allocated into two main groups ( $n = 30$ ) according to the type of sealer tested: **Group A** (AH Plus Jet) and **Group B** (Bio-C Sealer). Each group was then further divided into three subgroups ( $n = 10$  each) based on the dentine moisture condition, as follows:

1. **A1 and B1 (Dry):** Canals were dried by using sterile paper points followed by irrigation with 95% ethanol for 10 seconds and then re-dried with paper points.
2. **A2 and B2 (Normal moist):** Canals were blot dried by using sterile paper points until no visible moisture was detected [10].
3. **A3 and B3 (Wet):** Canals were left flooded with distilled water to simulate excessive dentine moisture [6].

### Root canal obturation

Obturation of the root canals was done by using the single-cone technique. A master gutta-percha cone corresponding to the final preparation size was coated with the assigned sealer and then inserted to the full working length. Specimens were subsequently stored at 37°C and 100% relative humidity for 7 days to ensure complete setting of the sealers.

### Specimen sectioning and preparation

Specimens were embedded in epoxy resin molds and then sectioned horizontally by using a slow-speed diamond saw under water cooling. Two sections of 2 mm thickness were obtained at 3 mm (apical level) and 8 mm (coronal level) from the apex of the roots. Sections were polished using silicon carbide papers (600-, 800-, and 1200-grit), followed by 0.05  $\mu$ m alumina suspension. Specimens were ultrasonically cleaned in distilled water for 3 minutes and then air dried.

### Scanning electron microscopy analysis

The specimens were mounted on aluminum stubs and sputter-coated with gold. The dentine–sealer interface was evaluated using a scanning electron microscope (Quanta 250 FEG; FEI, Netherlands) operating in backscattered electron mode at 15 kV. Interfacial adaptation was assessed qualitatively utilizing SEM micrographs by recording the presence of interfacial gaps and evaluating the continuity of the sealer–dentine interface. Images were obtained at a standardized magnification of  $\times 1200$ . The assessment was descriptive in nature based on a comparative observation between the tested groups.

**Table 1.** Classification and chemical composition of the investigated sealers.

| Sealer       | Manufacturer                             | Classification                                               | Chemical composition                                                                                                                                                                                                                                       |
|--------------|------------------------------------------|--------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| AH Plus Jet  | Dentsply Sirona; Ballaigues; Switzerland | Epoxy resin-based root canal sealer                          | Paste A: Bisphenol-A epoxy resin, calcium tungstate, bispheno-F epoxy resin, silica, zirconium oxide, iron oxide pigments.<br>Paste B: Dibenzyl diamine, calcium tungstate, tricyclodecane-diamine, aminoadamantane, silica, zirconium oxide silicone oil. |
| Bio-C Sealer | Angelus; Londrina; PR, Brazil            | Premixed calcium silicate-based bioceramic root canal sealer | Calcium silicates, zirconium oxide, silicon dioxide, calcium oxide, iron oxide, calcium aluminate, dispersing agents, and thickening agents.                                                                                                               |

### Energy-dispersive X-ray spectroscopy analysis

Elemental detection was conducted using the EDX system (Oxford Instruments, UK) attached to the SEM [18]. Three standardized regions were analyzed: the sealer–dentine interface, dentine side, and sealer side.

Weight (Wt%) and atomic (At%) percentages of the following elements were recorded: carbon (C), oxygen (O), silicon (Si), phosphorus (P), and calcium (Ca).

### Statistical analysis

Data were presented as mean  $\pm$  standard deviation (SD). Statistical analysis was carried out by using IBM SPSS Statistics version 28.0 (IBM Corp., Armonk, NY, USA). The effects of sealer type and dentine moisture conditions were assessed utilizing two-way ANOVA followed by Tukey's post hoc test for multiple comparisons when required. The level of significance was established at  $p < 0.05$ .

## Results

### Interfacial adaptation (SEM analysis)

In Figures 1–3, SEM analysis demonstrated variations in interfacial adaptation among all tested groups. The AH Plus Jet sealer showed a more continuous sealer–dentine interface with fewer visible gaps under dry conditions (A1), while more defects (disruptions) were recorded under wet conditions (A3).

Bio-C Sealer recorded the most favorable interfacial integrity under normal moist conditions (B2), showing intimate contact with the dentine walls and fewer detected defects (interfacial gaps) when compared with the dry and wet conditions.

Dentine moisture significantly affected the interfacial integrity at both the apical and coronal segments. The AH Plus Jet Sealer demonstrated superior adaptation with minimal interfacial gaps under dry conditions (A1), while wet conditions (A3) exhibited the least favorable adaptation with increased gap formation, and the moist condition (A2) recorded intermediate performance.

Figure 1 demonstrated that Bio-C Sealer recorded optimal interfacial adaptation with intimate sealer–dentine contact and reduced interfacial defects (discontinuities) under moist conditions (B2). Under dry (B1) and wet (B3) conditions, an inferior interfacial adaptation was observed although wet conditions showed a slight deterioration irrespective of the root level.

### Elemental analysis

The quantitative elemental data are presented in Tables 2–4. For both sealers, EDX spectra showed distinct elemental profiles. Bio-C Sealer recorded higher calcium and phosphorus signals, while AH Plus Jet showed predominant silicon and carbon peaks (Figures 2 and 3). The results of Tukey's post hoc tests are presented in Table 4.

### Carbon (C)

The AH Plus Sealer recorded a significantly higher carbon content when compared with Bio-C Sealer under all moisture conditions. At the apical level, carbon (wt%) in AH Plus varied from  $85.5 \pm 0.9$  (A1) to  $72.5 \pm 0.5$  (A3), while Bio-C Sealer showed lower values, in particular under normal moist conditions,  $17.5 \pm 0.5$  (B2) and  $29.2 \pm 0.2$  (B3). The statistical analysis indicated significant differences between respective subgroups, primarily A2 compared to B2 and A3 compared to B3 ( $p < 0.001$ ), as recorded in Table 3.

### Oxygen (O)

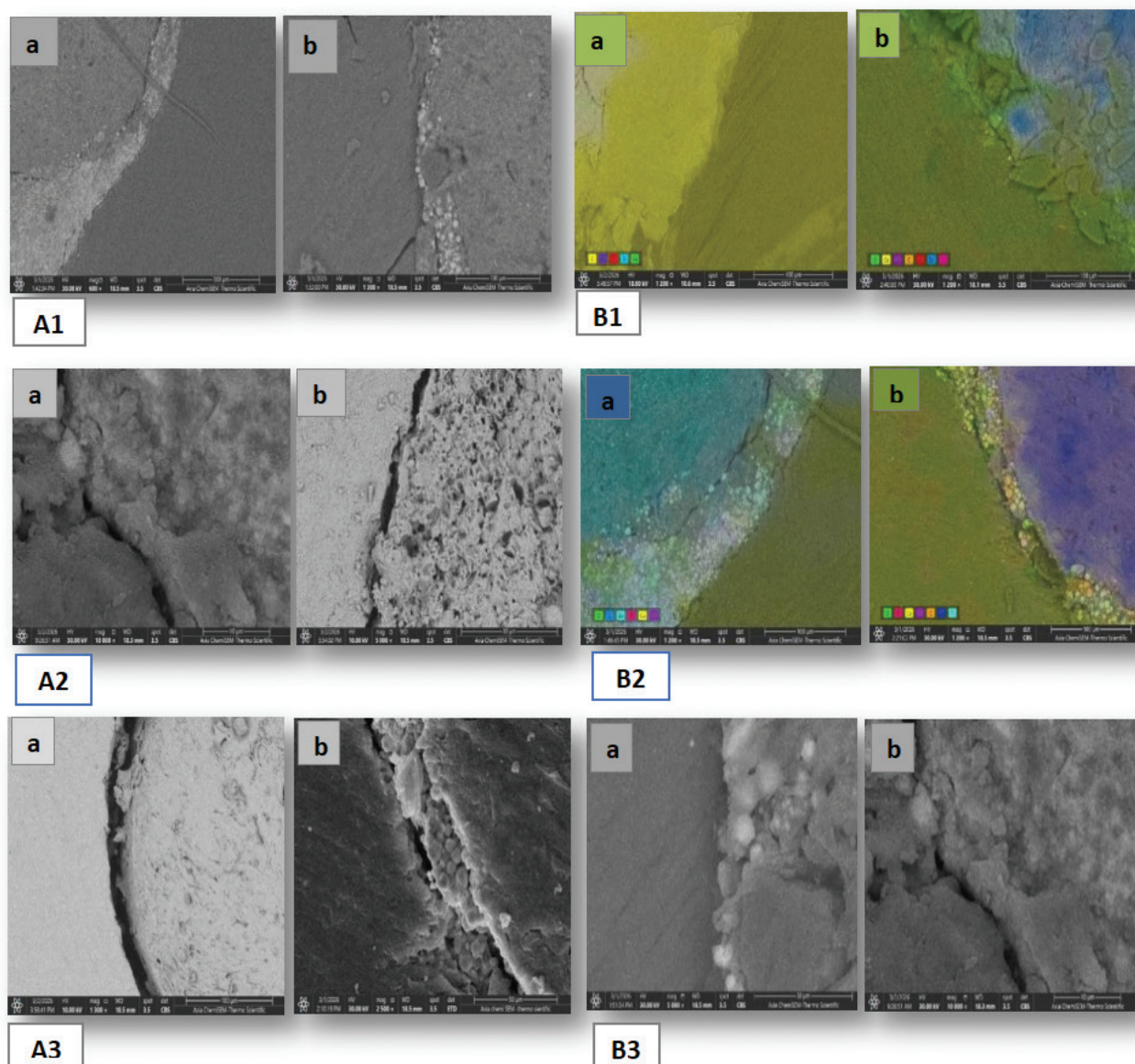
In Bio-C Sealer, the oxygen (O) levels were significantly higher compared with AH Plus. Bio-C Sealer recorded values of  $35.9 \pm 0.9$  (B2) and  $27.7 \pm 1.5$  (B3), while the AH Plus Sealer values ranging from  $8.7 \pm 0.1$  (A1) to  $21.2 \pm 0.2$  (A3). The differences were statistically significant for all study groups ( $p < 0.001$ ) as shown in Table 3.

### Silicon (Si)

AH Plus recorded a significantly higher silicon content. At the apical level, recorded values were  $9.2 \pm 0.2$  (A2) and  $4.60 \pm 0.4$  (A1), while lower values were recorded for Bio-C Sealer:  $3.0 \pm 0.25$  (B2) and  $1.7 \pm 0.8$  (B3). Statistically significant differences were demonstrated for most tested subgroups ( $p < 0.01$ ) (Table 3).

### Calcium (Ca)

Bio-C Sealer demonstrated significantly higher levels of calcium when compared with that of AH Plus Jet. At the apical level: Bio-C Sealer recorded values of  $43.6 \pm 0.65$  (B2) and  $41.4 \pm 0.4$  (B3), while the AH Plus Jet recorded values of  $1.30 \pm 0.05$  (A1) to  $10.6 \pm 0.4$  (A2). The differences were highly significant ( $p < 0.001$ ) (Table 3).



**Figure 1.** Representative SEM micrographs highlighting the sealer–dentine interface under three different dentine moisture conditions. A1–A3, respectively, show AH Plus under dry, normal moist, and wet conditions; B1–B3 represent Bio-C Sealer under the dry, moist, and wet conditions. Images marked (a) indicate the apical level and images marked and (b) indicate the coronal level. Dry-condition AH Plus specimens revealed fewer interfacial discontinuities, while the wet condition specimens recorded more visible gaps. Bio-C Sealer revealed the most favorable interfacial adaptation with the normal moist conditions. All micrographs were recorded at  $\times 1200$  magnification. SEM: scanning electron microscopy.

### Phosphorus (P)

Phosphorus was observed mainly in Bio-C Sealer, with higher values recorded under moist conditions. Statistically significant differences were demonstrated between tested groups, especially at the coronal level ( $p < 0.05$ ) (Table 3).

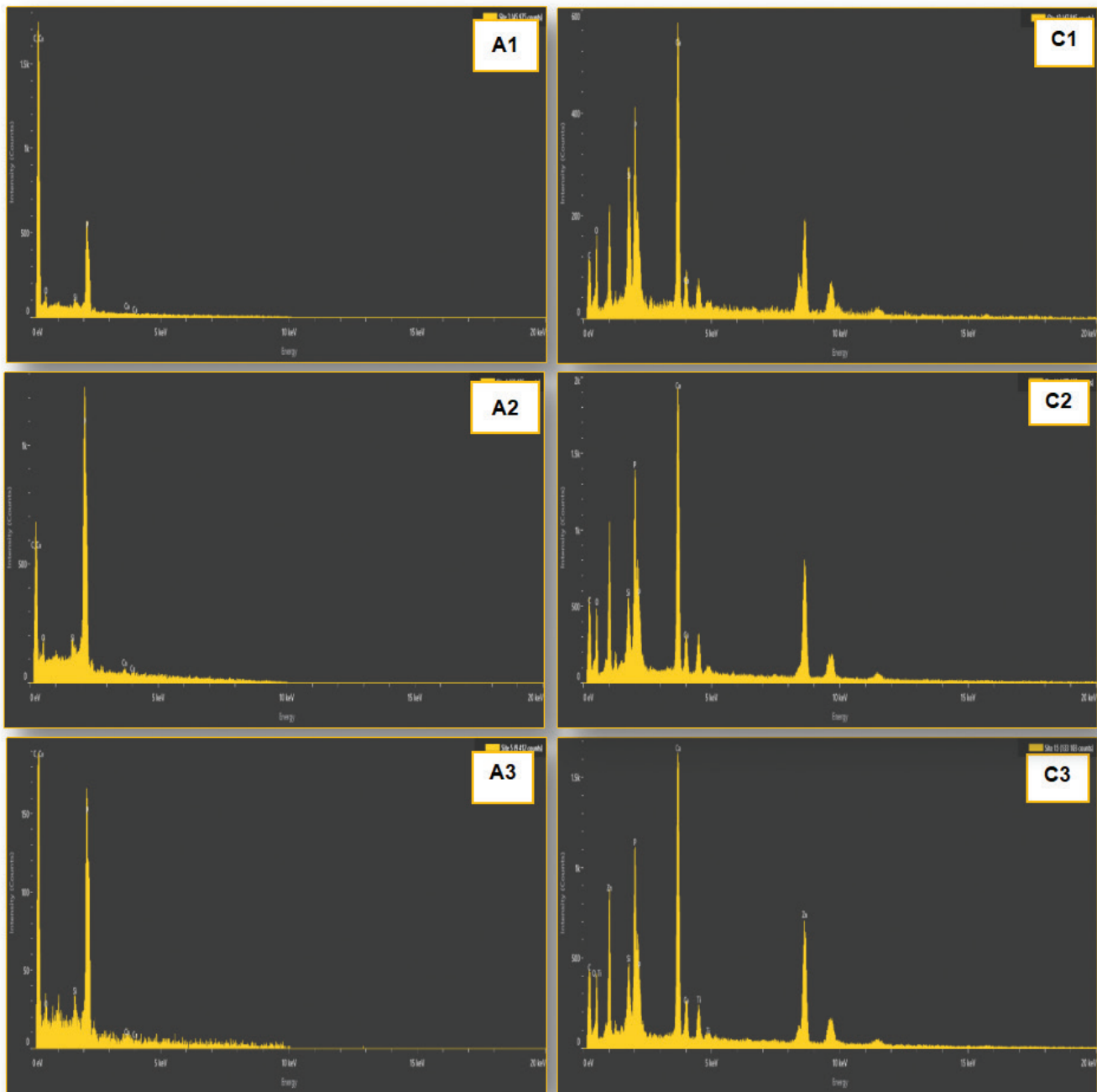
### Impact of root level

Elemental distribution revealed variation between the evaluated apical and coronal levels. In Bio-C Sealer, the levels of calcium were significantly higher at the apical segment when compared with the coronal one ( $p = 0.016$ ). On the other hand, no consistent regional differences in elemental composition were detected with AH Plus Jet.

### Discussion

This current investigation assessed the impact of dentine moisture on the interfacial integrity of a calcium silicate-based sealer compared with that of an epoxy resin-based sealer using SEM and EDX. Both hypotheses were rejected, highlighting that the type of sealer and dentine moisture condition significantly affected the outcomes. The results revealed that dentine moisture is a crucial factor affecting the interfacial performance of sealers, reflecting their various setting mechanisms and physicochemical properties [1, 18].

The single cone obturation technique was applied in this study to standardize the obturation procedure for all tested groups and to limit potential interfering factors associated with the compaction forces in addition to operator-related variables.



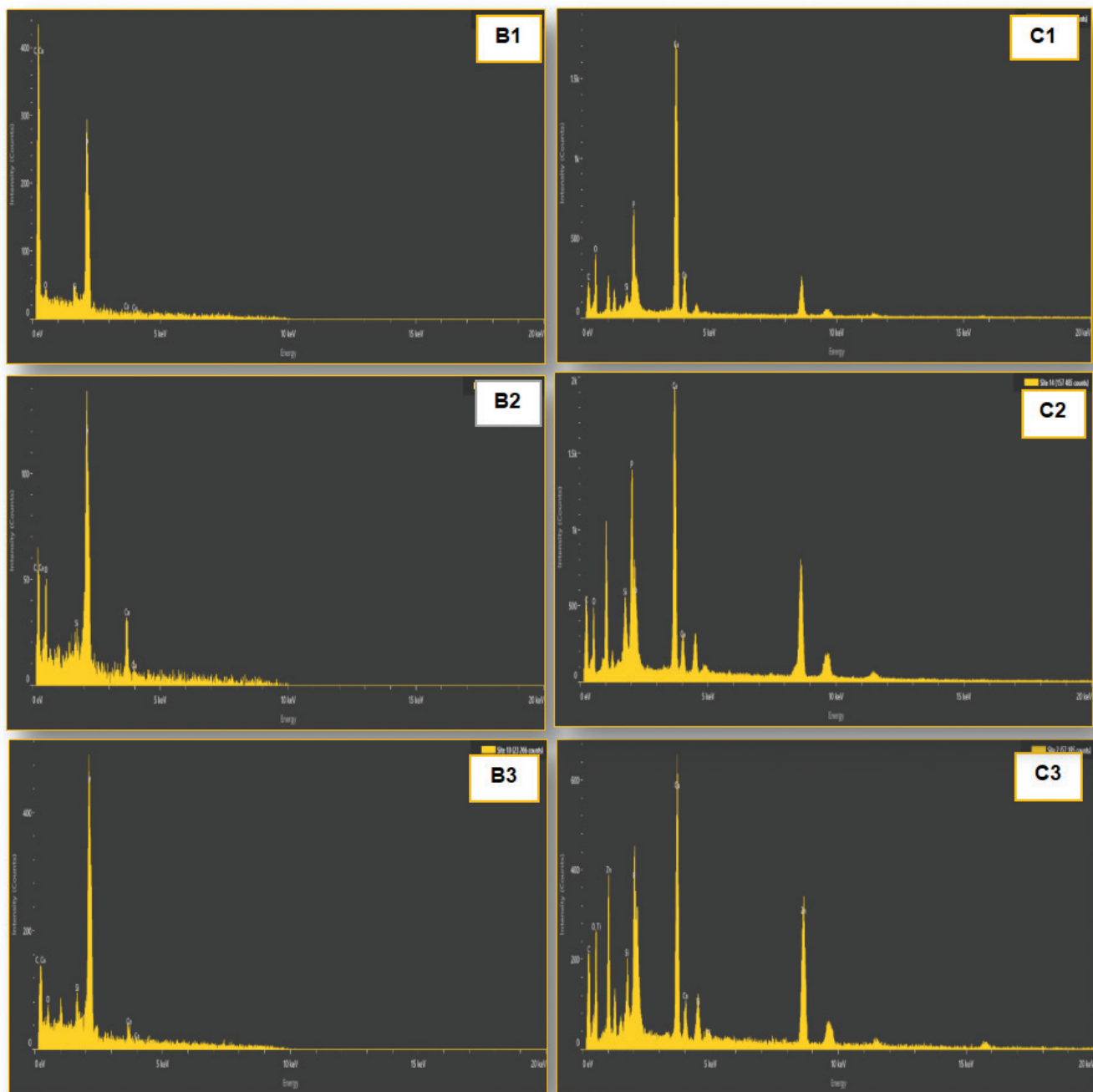
**Figure 2.** Representative EDX spectra of AH Plus under diverse moisture conditions: (A1) dry, (A2) normal moisture, and (A3) wet at the apical level, along with corresponding spectra at the coronal level (C1–C3), demonstrating the elemental composition across conditions. EDX: energy-dispersive X-ray spectroscopy analysis.

Although clinically, AH Plus Jet is mainly used with compaction-based obturation techniques, using the exact obturation procedure in all study groups ensured a more reliable comparison of the impact of dentine moisture conditions on the interfacial integrity. Thus, any recorded variations could be more confidently related to the tested variables rather than the obturation technique.

In addition to the intrinsic properties of sealers, some methodological factors may have an impact on the observed findings. The obturation technique could affect the distribution

of the root canal sealer and interface formation, especially when different materials exhibit distinct flow and setting behaviors.

The thickness of sealer may affect the setting mechanisms, interfacial integrity and dimensional stability, especially in materials that depend on hydration reactions. The storage conditions should also be considered, as temperature and humidity may affect the physicochemical behavior and the hydration process of calcium silicate-based root canal sealers [19]. These factors may partially explain the variations recorded between the tested sealers under different dentine moisture conditions.



**Figure 3.** Representative EDX spectra of Bio-C Sealer under diverse moisture conditions: (B1) dry, (B2) normal moisture, and (B3) wet at the apical level, along with corresponding spectra at the coronal level (C1–C3), demonstrating the elemental composition across conditions. EDX: energy-dispersive X-ray spectroscopy analysis.

Enhanced interfacial adaptation of AH Plus Jet root canal sealer under dry conditions can be ascribed to its hydrophobic epoxy resin matrix and moisture-sensitive polymerization reaction [1, 2]. Epoxy resin-based sealers undergo an epoxide-amine reaction that is water independent; however, the presence of residual moisture may interfere with polymer chain formation, reduce sealer penetration into the dentinal tubules, and may compromise interfacial integrity [19]. These findings are consistent with previous studies that reported improved bonding and marginal adaptation of AH Plus Jet in adequately

dried canals [4, 5]. Conversely, increased moisture has been associated with compromised adhesion and greater interfacial gap formation [20, 21].

On the other hand, Bio-C Sealer revealed optimal interfacial adaptation under moist conditions, which is consistent with its hydration-dependent setting reaction [22, 23]. Calcium silicate-based sealers require moisture to initiate hydration reactions, leading to the formation of calcium silicate hydrate (C-S-H) and calcium hydroxide. These products play an essential role in dimensional stability and interfacial continuity through micro-

**Table 2.** Mean  $\pm$  standard deviation (SD) of elemental composition (Wt% and At%) at the apical level for all subgroups.

| Elements  | A1 Dry          |                 | A2 Moist       |                | A3 Wet         |                | B1 Dry         |              | B2 Moist        |                 | B3 Wet         |                |
|-----------|-----------------|-----------------|----------------|----------------|----------------|----------------|----------------|--------------|-----------------|-----------------|----------------|----------------|
|           | Wt% $\pm$ SD    | At% $\pm$ SD    | Wt% $\pm$ SD   | At% $\pm$ SD   | Wt% $\pm$ SD   | At% $\pm$ SD   | Wt% $\pm$ SD   | At% $\pm$ SD | Wt% $\pm$ SD    | At% $\pm$ SD    | Wt% $\pm$ SD   | At% $\pm$ SD   |
| <b>C</b>  | 85.5 $\pm$ 0.9  | 90.60 $\pm$ 0.6 | 60.2 $\pm$ 0.2 | 73.2 $\pm$ 0.2 | 72.5 $\pm$ 0.5 | 80.1 $\pm$ 1.1 | 85.9 $\pm$ 0.4 | 89 $\pm$ 1.0 | 17.5 $\pm$ 0.5  | 29.7 $\pm$ 1.5  | 29.2 $\pm$ 0.2 | 46.2 $\pm$ 0.2 |
| <b>O</b>  | 8.7 $\pm$ 0.1   | 6.90 $\pm$ 1.0  | 19.9 $\pm$ 0.1 | 18.2 $\pm$ 0.2 | 21.2 $\pm$ 0.2 | 17.6 $\pm$ 0.6 | 14.1 $\pm$ 2.4 | 11 $\pm$ 0.9 | 35.9 $\pm$ 0.9  | 45.8 $\pm$ 1.8  | 27.7 $\pm$ 1.5 | 32.9 $\pm$ 2.5 |
| <b>Si</b> | 4.60 $\pm$ 0.4  | 2.10 $\pm$ 0.1  | 9.2 $\pm$ 0.2  | 4.8 $\pm$ 0.25 | 1.5 $\pm$ 0.5  | 0.7 $\pm$ 0.05 | -              | -            | 3 $\pm$ 0.25    | 2.2 $\pm$ 0.2   | 1.7 $\pm$ 0.8  | 1.2 $\pm$ 0.2  |
| <b>P</b>  | Not detected    | -               | -              | -              | -              | -              | -              | -            | -               | -               | -              | -              |
| <b>Ca</b> | 1.30 $\pm$ 0.05 | 0.40 $\pm$ 0.5  | 10.6 $\pm$ 0.4 | 3.9 $\pm$ 0.1  | 4.8 $\pm$ 0.2  | 1.6 $\pm$ 0.2  | -              | -            | 43.6 $\pm$ 0.65 | 22.2 $\pm$ 0.25 | 41.4 $\pm$ 0.4 | 19.7 $\pm$ 1.2 |

A1 = AH Plus (dry condition), A2 = AH Plus (normal moist condition), A3 = AH Plus (wet condition).

B1 = Bio-C Sealer (dry condition), B2 = Bio-C Sealer (normal moist condition), B3 = Bio-C Sealer (wet condition).

mechanical interlocking and chemical interaction with dentine [22, 24]. Inadequate moisture may limit hydration and impair setting, while excessive moisture may dilute the material and disrupt its microstructure, explaining the reduced adaptation observed under over-wet conditions [25, 26].

The EDX findings further support the bioactive properties of Bio-C Sealer. The significantly higher calcium and phosphorus levels recorded in this group, indicating an enhanced potential for calcium phosphate deposition at the sealer–dentine interface. This is in agreement with previous studies recording that calcium silicate-based materials release calcium ions that interact with phosphate ions from the surrounding environment, leading to the precipitation of apatite-like structures [26–29]. These findings are in agreement with a previous study demonstrating that bioceramic sealers exhibit favorable biocompatibility and bioactivity, primarily attributed to calcium hydroxide formation and their alkaline pH, which may enhance mineralized tissue formation in spite of the initial inflammatory response [30].

The formation of such mineralized interfacial layers may enhance sealing ability and contribute to long-term interfacial stability. These findings are consistent with other studies recording that the hydration behavior and calcium ion release from calcium silicate-based sealers can be significantly affected

by the surrounding chemical environment, which in turn modulates their bioactivity and interfacial performance [31].

The detected higher oxygen signals in Bio-C Sealer are likely associated with the hydration products and ongoing chemical reactions within this material, while the higher carbon levels in AH Plus Jet indicate its organic resin-based composition.

Calcium was also observed in AH Plus Jet, and this may be related to the interaction volume of EDX analysis, which includes the underlying dentine that is naturally rich in calcium. Assuming that the EDX analysis reflects the elemental composition of the interaction volume rather than the sealer alone, these findings should be considered with caution [1, 18].

Variations along the root canal levels were also examined, with obviously greater calcium signals recorded at the apical region in the Bio-C Sealer group. This may be related to the structural variations in dentine, including differences in density and permeability of the tubules, which may affect diffusion of ions and material tissue interaction [30, 32, 33]. The narrower anatomy and reduced diameter of the dentinal tubules at the apical region may enhance localized ion accumulation and promote mineral deposition. In contrast, the lack of obvious regional variations in the AH Plus group is consistent with its non-bioactive nature and limited ionic exchange [29].

**Table 3.** Independent samples *t*-test comparing elemental composition between AH Plus and Bio-C Sealer at apical and coronal levels.

| Element | Subgroups | Apical segment |          |           |          | Coronal segment |          |           |          |
|---------|-----------|----------------|----------|-----------|----------|-----------------|----------|-----------|----------|
|         |           | Weight %       |          | Atomic %  |          | Weight %        |          | Atomic %  |          |
|         |           | Mean diff      | <i>P</i> | Mean diff | <i>P</i> | Mean diff       | <i>P</i> | Mean diff | <i>P</i> |
| C       | A1vsB1    | -0.4           | 0.628    | 1.6       | 0.076    | 6.4             | 0.001*   | 10.1      | 0.000*   |
|         | A2vsB2    | 42.7           | 0.00*    | 43.5      | 0.00*    | 6.8             | 0.111    | 10.5      | 0.004*   |
|         | A3vsB3    | 43.3           | 0.00*    | 33.9      | 0.00*    | 1.6             | 0.012*   | 3.2       | 0.062    |
| O       | A1vsB1    | -5.4           | 0.00*    | -4.1      | 0.006*   | -7.1            | 0.001*   | -10.6     | 0.000*   |
|         | A2vsB2    | -16            | 0.01*    | -27.6     | 0.00*    | -4.9            | 0.00*    | -8.1      | 0.000*   |
|         | A3vsB3    | -6.5           | 0.00*    | -15.3     | 0.00*    | -4.6            | 0.00*    | -5.8      | 0.000*   |
| Si      | A1vsB1    | 4.6            | 0.00*    | 2.1       | 0.006*   | 6.5             | 0.00*    | 4.7       | 0.001*   |
|         | A2vsB2    | 6.2            | 0.00*    | 2.6       | 0.00*    | 4.2             | 0.276    | 1.6       | 0.014*   |
|         | A3vsB3    | -0.2           | 0.708    | -0.5      | 0.014*   | 0.6             | 0.37     | 0.5       | 0.30     |
| P       | A1vsB1    | -              | -        | -         | -        | -0.8            | 0.217    | -0.7      | 0.232    |
|         | A2vsB2    | -              | -        | -         | -        | 4.6             | 0.022    | 3.1       | 0.106    |
|         | A3vsB3    | -              | -        | -         | -        | 1.1             | 0.152    | 0.8       | 0.012*   |
| Ca      | A1vsB1    | 1.3            | 0.00*    | 0.4       | 0.005*   | -6.2            | 0.00*    | -4.7      | 0.005*   |
|         | A2vsB2    | -33            | 0.00*    | -18.3     | 0.00*    | 13.2            | 0.00*    | 6.4       | 0.000*   |
|         | A3vsB3    | -36.6          | 0.00*    | -18.1     | 0.00*    | 4.8             | 0.001*   | 2.6       | 0.004*   |

\*Significant difference ( $p \leq 0.05$ ). **A1:** AH Plus in dry condition. **A2:** AH Plus in normal moist. **A3:** AH Plus in wet condition. **B1:** Bio-C Sealer in dry condition. **B2:** Bio-C Sealer in normal moist. **B3:** Bio-C Sealer in wet condition.

**Table 4.** Tukey's post hoc test for multiple comparisons of elemental composition among different dentine moisture conditions.

| Element | Apically  |           |        |           |        | Coronally |           |        |           |        |
|---------|-----------|-----------|--------|-----------|--------|-----------|-----------|--------|-----------|--------|
|         | Subgroups | Weight %  |        | Atomic %  |        | Subgroups | Weight %  |        | Atomic %  |        |
|         |           | Mean diff | P      | Mean diff | P      |           | Mean diff | P      | Mean diff | P      |
| C       | A1vsA2    | -3.8      | 0.006* | -7.7      | 0.337  | B1vs B2   | -3.4      | 0.00*  | -7.3      | 0.836  |
|         | A1vsA3    | -4.1      | 0.00*  | -8.2      | 0.003* | B1vsB3    | -8.9      | 0.623  | -15.1     | 0.00*  |
|         | A2vsB3    | -0.3      | 1.0    | -0.5      | 0.264  | B2vs B3   | -5.5      | 0.029* | -7.8      | 0.044* |
| O       | A1vsA2    | 5.6       | 0.036* | 6.5       | 0.005* | B1vs B2   | 7.8       | 0.00*  | 9         | 0.01   |
|         | A1vsA3    | 6.1       | 0.00*  | 7.1       | 0.003* | B1vsB3    | 8.6       | 0.00*  | 11.9      | 0.00*  |
|         | A2vsB3    | 0.5       | 1.0    | 0.6       | 0.264  | B2vs B3   | 0.8       | 0.065  | 2.9       | 0.044* |
| Si      | A1vsA2    | 4.2       | 0.903  | 3         | 0.00*  | B1vs B2   | 0.3       | 0.00*  | 0.3       | 0.002* |
|         | A1vsA3    | 4.4       | 0.870  | 3.1       | 0.00*  | B1vsB3    | -1.5      | 0.00*  | -1.1      | 0.00*  |
|         | A2vsB3    | 0.2       | 0.82   | 0.1       | 0.066  | B2vs B3   | -1.8      | 0.507  | -1.4      | 0.005* |
| P       | A1vsA2    | 0.3       | 0.00*  | 0.1       | 0.00*  | B1vs B2   | 3.1       | 0.004* | 2.1       | 0.120  |
|         | A1vsA3    | 0         | 0.00*  | -0.1      | 0.00*  | B1vsB3    | 1.9       | 0.00*  | 1.4       | 0.00*  |
|         | A2vsB3    | -0.3      | 0.943  | -0.2      | 0.174  | B2vs B3   | -1.2      | 0.011* | -0.7      | 0.028* |
| Ca      | A1vsA2    | 4.5       | 0.511  | 2         | 0.00*  | B1vs B2   | 1.4       | 0.00*  | 0.2       | 0.049  |
|         | A1vsA3    | 3.8       | 0.87   | 1.7       | 0.00*  | B1vsB3    | 14.8      | 0.00*  | 7         | 0.00*  |
|         | A2vsB3    | -0.7      | 0.997  | -0.3      | 0.569  | B2vs B3   | 13.4      | 0.01*  | 7.8       | 0.168  |

\*Significant difference ( $p \leq 0.05$ ). **A1**: AH Plus in dry condition. **A2**: AH Plus in normal moist. **A3**: AH Plus in wet condition. **B1**: Bio-C Sealer in dry condition. **B2**: Bio-C Sealer in normal moist. **B3**: Bio-C Sealer in wet condition.

The findings of the current investigation are in partial agreement with those recorded by Pelozo et al. [34], who found that dentine moisture conditions affected the adhesive properties of calcium silicate-based root canal sealers. While their study primarily examined the bonding strength and adhesive properties of the interface, the present investigation assessed the interfacial integrity and elemental composition by using (SEM/EDX) analyses and integrated a comparison with an epoxy resin-based (AH Plus Jet) root canal sealer. The enhanced performance of Bio-C Sealer under moist conditions observed in both studies supports the hypothesis that calcium silicate-based root canal sealers perform optimally with residual dentine moisture due to their hydration-dependent setting reactions.

Clinically, these findings highlight the importance of tailoring canal drying protocols in accordance with the type of sealer. Over-dryness may compromise the clinical outcomes of calcium silicate-based sealers, while residual moisture may negatively influence the performance of epoxy resin-based materials. Thus, attaining an optimal residual moisture condition is essential to enhance the clinical performance and treatment outcomes.

### Limitations

1. The in vitro design may not fully replicate the clinical settings, where factors like tissue fluids, microbial activity, and functional loading may affect the sealer performance. Additionally, mechanical properties such as push out bonding strength and micro-leakage were not evaluated. Further studies should integrate clinically representative models and investigate the long-term performance of these materials.
2. The single-cone obturation protocol was applied for all tested groups in order to standardize the experimental conditions. While this approach facilitated a direct comparison between the sealers, it may not reflect the clinical use of AH

Plus Jet, which is more commonly applied with compaction-based obturation techniques. Consequently, the thickness and volume of sealer all around the gutta-percha cone may have affected the observed interfacial properties. Additionally, the interfacial integrity was assessed qualitatively by using SEM rather than quantitative assessment of gap formation; thus, conclusions about interface quality should be interpreted with caution. Future studies should include quantitative interfacial analyses, bonding strength evaluation, and long-term investigations under more representative clinical conditions.

### Conclusions

Dentinal moisture conditions significantly affect the interfacial adaptation and elemental characteristics of root canal sealers. AH Plus Jet recorded optimal performance under dry conditions, while the Bio-C Sealer demonstrated improved adaptation and bioactivity under moist conditions. The increased calcium and phosphorus levels recorded in the Bio-C group support its bioactive potential and its ability to promote interfacial mineralization.

### Declaration of interest

The authors declare that they have no known competing personal relationships or financial interests that could have appeared to affect the work reported in this manuscript.

### Ethical approval

Ethical approval was obtained from the Research Ethics Committee, College of Dentistry, Mustansiriyah University, Baghdad, Iraq (Approval No MUOPR29). All human extracted teeth used in this work were collected and handled in

accordance with the institutional ethical guidelines and relevant regulations.

### Conflicts of interest

The authors of this article declare that they have no conflicts of interest.

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### Data availability statement

The data supporting the findings of this research are available from the authors upon reasonable request.

### Author contributions

Maha M. Yahya contributed to conceptualization, methodology, formal analysis, data curation, and writing the original draft. Baidaa Mohammed Zeidan contributed to methodology, formal analysis, and investigation. Njwan Fadhel Shehab contributed to investigation, manuscript review, and editing. All authors read and approved the final version of the manuscript.

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### References

- [1] Błaszczuk-Pośpiech A, Struzik N, Szymonowicz M, Sareło P, Wiśniewska-Wrona M, Wiśniewska K, et al. Endodontic sealers and innovations to enhance their properties: a current review. *Materials* (Basel). 2025;18(18):4259. <https://doi.org/10.3390/ma18184259>
- [2] Kim M, Hayashi M, Yu B, Lee TK, Kim RH, Jo DW. Cytotoxicity and genotoxicity of epoxy resin-based root canal sealers before and after setting procedures. *Life* (Basel). 2022;12(6):847. <https://doi.org/10.3390/life12060847>
- [3] Malta CP, Barcelos RCS, Fernandes PS, Martins MO, Sagrillo MR, Bier CAS, et al. In silico toxicity and immunological interactions of components of calcium silicate-based and epoxy resin-based endodontic sealers. *Clin Oral Investig*. 2024;28(2):148. <https://doi.org/10.1007/s00784-024-05548-y>
- [4] Razmi H, Bolhari B, Dashti NK, Fazlyab M. The effect of canal dryness on bond strength of bioceramic and epoxy-resin sealers after irrigation with sodium hypochlorite or chlorhexidine. *Iran Endod J*. 2016;11(2):129–33. <https://doi.org/10.7508/iej.2016.02.011>
- [5] Piazza B, Rivera-Peña ME, Alcalde MP, de Vasconcelos BC, Duarte MAH, de Moraes IG, et al. The influence of humidity on intra-tubular penetration and bond strength of AH plus and MTA fillapex: an in vitro study. *Eur Endod J*. 2017;3(1):48–54. <https://doi.org/10.5152/eej.2017.17036>
- [6] Kadı G, Özlek E, Saed YMK. Effect of different drying protocols on the bond strength of a bioceramic root canal sealer. *Aust Endod J*. 2026;52(1):70–6. <https://doi.org/10.1111/aej.70032>
- [7] Guivarc'h M, Jeanneau C, Giraud T, Pommel L, About I, Azim AA, et al. An international survey on the use of calcium silicate-based sealers in non-surgical endodontic treatment. *Clin Oral Investig*. 2020;24(1):417–24. <https://doi.org/10.1007/s00784-019-02920-1>
- [8] Tolosa-Monfà A, Veroni A, Blasi-Cabús J, Ballester-Palacios ML, Berástegui-Jimeno E. Cytotoxicity comparison of Bio-C sealer against multiple root canal sealers. *J Clin Exp Dent*. 2023;15(2):e110–17. <https://doi.org/10.4317/jced.59868>
- [9] Voegeli G, Di Bella E, Mekki M, Machtou P, Bouillaguet S. Effect of a modified irrigation protocol on the cleanliness of moderately curved canals. *Eur J Dent*. 2020;15(1):90–5. <https://doi.org/10.1055/s-0040-1715985>
- [10] Wang JS, Bai W, Wang Y, Liang YH. Effect of different dentin moisture on the push-out strength of bioceramic root canal sealer. *J Dent Sci*. 2023;18(1):129–34. <https://doi.org/10.1016/j.jds.2022.06.025>
- [11] Marta M, Chivu OR, Marian D, Enache I, Veja I, Pitic DE, et al. Elemental composition and dentin bioactivity at the interface with ah plus bioceramic sealer: an energy-dispersive x-ray spectroscopy study. *Appl Sci*. 2024;14(24):11867. <https://doi.org/10.3390/app142411867>
- [12] Bhor S, Rao AS, Shah U, Mathur M, Reda R, Pagnoni F, et al. Comparative evaluation of the sealing ability of a bioceramic sealer (iroot sp) with AH plus sealer with root canal dentin using three different techniques of sealer application: a combined dye extraction and scanning electron microscope study. *J Compos Sci*. 2023;7:106. <https://doi.org/10.3390/jcs7030106>
- [13] Ha JH, Kim HC, Kim YK, Kwon TY. An evaluation of wetting and adhesion of three bioceramic root canal sealers to intraradicular human dentin. *Materials* (Basel). 2018;11(8):1286. <https://doi.org/10.3390/ma11081286>
- [14] Cabral MA, Limoeiro AG, De Martin AS, Fontana CE, Pelegri RA, Bueno CE, et al. Influence of root canal moisture conditions on the bond strength of endodontic sealers to dentin. *Res Soc Dev*. 2022;11(11):e285111133714. <https://doi.org/10.33448/rsd-v11i11.33714>
- [15] Mann NS, Mann NK, Kapur R. Evaluating the penetration efficacy of calcium silicate-based bioceramic sealers into dentinal tubules with cold lateral compaction technique using confocal laser scanning microscopy: an in vitro study. *J Conserv Dent Endod*. 2025;28(2):150–4. [https://doi.org/10.4103/JCDE.JCDE\\_754\\_24](https://doi.org/10.4103/JCDE.JCDE_754_24)
- [16] Margono A, Pratiwi RS, Nofarina AD, Npa DA, Maharti ID, Rossetti R. Biocompatibility and bioactivity evaluation of novel calcium silicate-based sealer: in vitro study on human dental pulp stem cells. *Eur J Dent*. 2025;19(3):777–83. <https://doi.org/10.1055/s-0045-1802566>
- [17] López-García S, Pecci-Lloret MR, Guerrero-Gironés J, Pecci-Lloret MP, Lozano A, Llena C, et al. Comparative cytocompatibility and mineralization potential of bio-c sealer and totalfill bc sealer. *Materials* (Basel). 2019;12(19):3087. <https://doi.org/10.3390/ma12193087>
- [18] Scimeca M, Bischetti S, Lamsira HK, Bonfiglio R, Bonanno E. Energy dispersive X-ray (EDX) microanalysis: a powerful tool in biomedical research and diagnosis. *Eur J Histochem*. 2018;62(1):2841. <https://doi.org/10.4081/ejh.2018.2841>
- [19] Sfeir G, Zogheib C, Patel S, Giraud T, Nagendrababu V, Bukiet F. Calcium silicate-based root canal sealers: a narrative review and clinical perspectives. *Materials* (Basel). 2021;14(14):3965. <https://doi.org/10.3390/ma14143965>
- [20] Zmener O, Pameijer CH, Serrano SA, Videira M, Macchi RL. Significance of moist root canal dentin with the use of methacrylate-based endodontic sealers: an in vitro coronal dye leakage study. *J Endod*. 2008;34(1):76–9. <https://doi.org/10.1016/j.joen.2007.10.012>
- [21] Dias KC, Soares CJ, Steier L, Versiani MA, Rached-Júnior FJ, Pécora JD, et al. Influence of drying protocol with isopropyl alcohol on the bond strength of resin-based sealers to the root dentin. *J Endod*. 2014;40(9):1454–8. <https://doi.org/10.1016/j.joen.2014.02.021>

- [22] Zordan-Bronzel CL, Torres FFE, Tanomaru-Filho M, Chávez-Andrade GM, Bosso-Martelo R, Guerreiro-Tanomaru JM. Evaluation of physicochemical properties of a new calcium silicate-based sealer, Bio-C sealer. *J Endod*. 2019;45(10):1248–52. <https://doi.org/10.1016/j.joen.2019.07.006>
- [23] Candeiro GT, Correia FC, Duarte MA, Ribeiro-Siqueira DC, Gavini G. Evaluation of radiopacity, pH, release of calcium ions, and flow of a bioceramic root canal sealer. *J Endod*. 2012;38(6):842–5. <https://doi.org/10.1016/j.joen.2012.02.029>
- [24] Watson TF, Atmeh AR, Sajini S, Cook RJ, Festy F. Present and future of glass-ionomers and calcium-silicate cements as bioactive materials in dentistry: biophotonics-based interfacial analyses in health and disease. *Dent Mater*. 2014;30(1):50–61. <https://doi.org/10.1016/j.dental.2013.08.202>
- [25] Al-Haddad AY, Kutty MG, Abu Kasim NH, Che Ab Aziz ZA. The effect of moisture conditions on the constitution of two bioceramic-based root canal sealers. *J Dent Sci*. 2017;12(4):340–6. <https://doi.org/10.1016/j.jds.2017.03.008>
- [26] Alsofi L. Bioactivity and element composition of three endodontic root canal sealers. *J Contemp Dent Pract*. 2025;26(1):62–70. <https://doi.org/10.5005/jp-journals-10024-3791>
- [27] Vargas AW, Pauletto G, Oliveira Pilecco R, Cechin LE, Pereira GKR, Morgental RD. Different root canal drying protocols for AH Plus bioceramic sealer: an in vitro study. *Eur J Oral Sci*. 2025;133(5):e70028. <https://doi.org/10.1111/eos.70028>
- [28] Zamparini F, Prati C, Taddei P, Spinelli A, Di Foggia M, Gandolfi MG. Chemical-physical properties and bioactivity of new premixed calcium silicate-bioceramic root canal sealers. *Int J Mol Sci*. 2022;23:13914. <https://doi.org/10.3390/ijms232213914>
- [29] Sheela S, Nassar M, AlGhalban FM, Gorduysus MO. In vitro cytotoxicity and mineralization potential of an endodontic bioceramic material. *Eur J Dent*. 2023;17(2):548–55. <https://doi.org/10.1055/s-0042-1750778>
- [30] Inada RNH, Silva ECA, Lopes CS, Queiroz MB, Torres FFE, da Silva GF, et al. Biocompatibility, bioactivity, porosity, and sealer/dentin interface of bioceramic ready-to-use sealers using a dentin-tube model. *Sci Rep*. 2024;14(1):16768. <https://doi.org/10.1038/s41598-024-66616-7>
- [31] Divya PM, Jena A, Mohanty S, Shashirekha G, Mallick RR, Sarangi P. Influence of irrigating solutions on the hydration of calcium silicate-based dental biomaterials: an in vitro study. *J Conserv Dent Endod*. 2025;28(8):758–63. [https://doi.org/10.4103/JCDE.JCDE\\_335\\_25](https://doi.org/10.4103/JCDE.JCDE_335_25)
- [32] Mert DB, Gençoğlu N. Evaluation of the physical properties of different bioceramic-based root canal sealers. *Bezmialem Sci*. 2024;12(2):224–30. <https://doi.org/10.14235/bas.galenos.2024.84429>
- [33] Jeong JW, DeGraft-Johnson A, Dorn SO, Di Fiore PM. Dentinal tubule penetration of a calcium silicate-based root canal sealer with different obturation methods. *J Endod*. 2017;43(4):633–7. <https://doi.org/10.1016/j.joen.2016.11.023>
- [34] Pelozo LL, Souza-Gabriel AE, Alves Dos Santos GN, Camargo RV, Lopes-Olhê FC, Sousa-Neto MD, et al. Canal drying protocols to use with calcium silicate-based sealer: effect on bond strength and adhesive interface. *J Endod*. 2023;49(9):1154–60. <https://doi.org/10.1016/j.joen.2023.07.015>